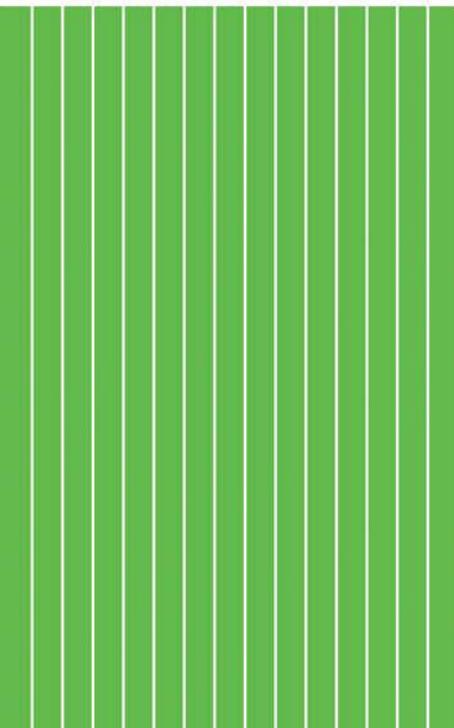


NORWAY

TRENDS AND SOURCES OF ZONOTIC AGENTS IN HUMANS, FOODSTUFFS ANIMALS AND FEEDINGSTUFF

- including information on foodborne outbreaks, antimicrobial resistance in zoonotic agents and some pathogenic microbiological agents

IN 2010



Norway

Trends and sources of zoonoses and zoonotic agents in humans, foodstuffs, animals and feedingstuffs

- including information on foodborne outbreaks, antimicrobial resistance in zoonotic agents and some pathogenic microbiological agents

In 2010

The Report referred to in Article 9 of Directive 2003/99/EC

Information on the reporting and monitoring system

Country: Norway
Reporting Year: 2010

Institutions and laboratories involved in reporting and monitoring:

The Norwegian Food Safety Authority (NFSA)

The Norwegian Food Safety Authority (NFSA) is the competent authority for the purpose of Directive 2003/99/EC of the European Parliament and of the Council.
Contributing with data and text.

The Norwegian Veterinary Institute (NVI)

The Norwegian Veterinary Institute (NVI) is a governmental agency funded by the Ministry of Agriculture and Food, Ministry of Fisheries and Coastal Affairs and the Norwegian Research Council. The primary function is the supply of independent research based advisory support to the governing authorities regarding animal health, fish health and food safety.
Contributing with data and text. The reporting officer is employed at the Zoonosis Centre at NVI.

The National Institute of Nutrition and Seafood Research (NIFES)

The National Institute of Nutrition and Seafood Research (NIFES) is a research institute with administrative tasks. The institute is linked directly to the Ministry of Fisheries and Coastal Affairs and act as an advisor to the Ministry in matters concerning the "fjord to fork" production chain of seafood (both wild and farmed). NIFES also provides independent and research based advisory support to other governmental bodies and to the Norwegian fisheries and aquaculture industries.
Contributing with data and text.

The Norwegian Institute of Public Health (NIPH)

The Norwegian Institute of Public Health (NIPH) is the national governmental centre for communicable disease prevention and control. The institute performs research and surveillance of communicable diseases in man and advises governmental and municipal authorities and the public on the prevention of communicable diseases, outbreaks and antimicrobial resistance. The institute also has responsibilities concerning chronic disease epidemiology, environmental medicine and forensic toxicology.
Contributing with data and text.

Suggested citation: Hofshagen M, Heier B T, Hauge K: Norway 2010 - Trends and source of zoonoses and zoonotic agents in humans foodstuffs, animals and feedingstuffs.

Preface

This report is submitted to the European Commission in accordance with Article 9 of Council Directive 2003/99/EC¹. The information has also been forwarded to the European Food Safety Authority (EFSA) and to the EFTA Surveillance Authority (ESA).

The report contains information on trends and sources of zoonoses and zoonotic agents in Norway during the year 2010. The information covers the occurrence of these diseases and agents in humans, animals, foodstuffs and for *Salmonella* also feedingstuffs. In addition the report includes data on antimicrobial resistance in some zoonotic agents and commensal bacteria as well as information on epidemiological investigations of foodborne outbreaks. Complementary data on susceptible animal populations in the country is also given. The information given covers both zoonoses that are important for the public health in the whole European Community as well as zoonoses, which are relevant on the basis of the national epidemiological situation.

The report describes the monitoring systems in place and the prevention and control strategies applied in the country. For some zoonoses this monitoring is based on legal requirements laid down by the Community Legislation, while for others national requirements are applied.

The report presents the results of the examinations carried out in 2010. A national evaluation of the epidemiological situation, with special reference to trends and sources of zoonotic infections, is given. Whenever possible, the relevance of findings in foodstuffs and animals to zoonoses cases in humans is evaluated.

The information covered by this report is used in the annual Community Summary Report on zoonoses that is published each year by EFSA.

¹Directive 2003/99/EC of the European Parliament and of the Council of 12 December 2003 on the monitoring of zoonoses and zoonotic agents, amending Decision 90/424/EEC and repealing Council Directive 92/117/EEC, OJ L 325, 17.11.2003, p. 31

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Animal populations

The relevance of the findings on zoonoses and zoonotic agents has to be related to the size and nature of the animal population in the country.

Sources of information

Data on herds and animals: Register of Production Subsidies.

Data on slaughtered animals: Register of Slaughtered Animals.

Dates the figures relate to and the content of the figures

Data on herds and animals: As of 31 July 2010.

Definitions used for different types of animals, herds, flocks and holdings as well as the types covered by the information

Herd means an animal or group of animals kept on a holding as an epidemiological unit (article 2.3(a) of Regulation (EC) No 2160/2003). In Norway, there is generally only one herd of the same animal species per holding.

A flock (poultry) is defined as all poultry of the same health status kept on the same premises or in the same enclosure and constituting a single epidemiological unit; in the case of housed poultry, this includes all birds sharing the same airspace (article 2.3(b) of Regulation (EC) No 2160/2003).

National evaluation of the numbers of susceptible population and trends in these figures

For cattle, swine, sheep, goat and poultry (layers and broilers) there has been a downward trend in the number of herds/holdings during the last decade. However, the average number of animals per herd/holding has increased.

Geographical distribution and size distribution of the herds, flocks and holdings

Cattle: Most of the cattle herds are dairy herds, the average herd size being 21.4 cows. There are also a number of specialized beef herds with an average number of suckling cows of 14.6. A few herds are combined dairy and beef herds. The cattle herds are distributed throughout Norway with the main part being in the western and middle parts of Norway.

Swine: The Norwegian swine population is relatively small with products destined for the national market. A national breeding program is organized by the industry. Approximately 120 approved elite and multiplier breeding herds house 11% of the live sows in the population, while more than 95% of the sows producing piglets for fattening and slaughter are raised in these herds. The swine population is denser in some counties and about 60% of the swine production is concentrated in four counties in the southern and middle part of Norway.

Sheep: The Norwegian sheep flocks are widely distributed over the country, with the largest population found in the southwest. The sheep population consists of combined meat and wool producing breeds, with various Norwegian breeds predominating.

Goat: The Norwegian goat population is principally composed of one Norwegian breed. The main product is milk used for cheese production. The goat flocks are located in some mountainous regions in the southern part of the country, in the fjord districts of the western part, and in the northern counties.

Poultry: The Norwegian poultry production has a hierarchical structure and is strictly regulated. Egg and broiler meat production are the most important branches, but the number of holdings keeping turkey and other species is increasing. The Norwegian layer population consists of two strains (Lohmann white and Shaver white). The layer population is located throughout Norway. The commercial broiler production consists of one strain (Ross). The broiler production is mainly located in five counties in the southern and middle part of Norway.

Additional information

The livestock production in Norway is targeted for the national market. Until 1999 there was a general ban on the import of live animals and animal products to Norway. Following the extension of the European Economic Area (EEA) Agreement 1 January 1999 regarding Veterinary and Phytosanitary matters, the general ban was lifted. However, imports of live animals remained limited.

Table Susceptible animal population

	Number of holdings (flocks) ¹	Livestock numbers (live animals) ¹	Number of slaughtered animals ²
Cattle			
Dairy cows and heifers	10,200	213,800	
Mixed herds	950	32,100	
Meat production animals	4,200	61,500	
In total	16,800	872,100	306,900
Deer³	80	7,100	
Gallus gallus (fowl)			
Grandparent breeding flocks – egg production line	2 (3)		
Parent breeding flocks – egg production line ⁴	11 (20)		
Parent breeding flocks – meat production line ⁴	68 (140)		
Laying hens ⁵	660 (988)	3,808,900	882,400
Broilers	600 (4600)		60,194,500
Goats			
Milk goats	400	36,900	
In total	1,300	67,600	24,300
Pigs			
Breeding animals	1,400	57,800	
Fattening pigs	2,200	461,400	
In total	2,400	846,700	1,565,700
Sheep			
Animals over 1 years	14,600	887,600	
In total	14,800	2,296,900	1,228,100
Solipeds – horses			1500
Turkeys, ducks and geese			
Parent breeding flocks ⁴	9 (17)		
In total	79 ⁶	363,200	1,295,700

Numbers between 100 and 1000 rounded to the nearest ten, numbers >1000 rounded to the nearest hundred.

¹ Register of Production Subsidies as of 31 July, 2010

² Register of Slaughtered Animals.

³ As of 31.07.2010

⁴ Only production flocks.

⁵ Only flocks >250 birds.

⁶ Only flocks >25 birds.

Salmonellosis

General evaluation of the national situation

History of the disease and/or infection in the country

The situation regarding *Salmonella* in feedingstuffs, animals and food produced in Norway has for many years been very good. Approximately 75-80% of the cases of salmonellosis in humans are acquired abroad.

National evaluation of the recent situation, the trends and sources of infection

There is no alarming development in the number of salmonellosis cases in humans, neither for domestic nor imported cases. However, there seem to have been a slightly increasing trend in domestic infections during the last decade.

For feedingstuffs and animals, the situation is very good and has been so for many years. Regarding food, the food produced in Norway is virtually free from *Salmonella*. Risk of exposure is mainly associated with international trade in food.

Relevance of the findings in animals, feedingstuffs and foodstuffs to human cases

The Norwegian *Salmonella* Control Programmes have documented that live cattle, swine, and poultry in Norway as well as domestically produced food products of animal origin are virtually free from *Salmonella*. Each year, approximately 75-80% of reported cases of salmonellosis in humans have acquired the infection abroad. This illustrates that domestic food products of animal origin represent a small risk to the consumer in regard to *Salmonella*, an assumption that is supported by case control studies.

Salmonellosis in humans

Reporting system in place for the human cases

Human cases are reported to the Norwegian Surveillance System for Communicable Diseases (MSIS), from microbiological laboratories as well as from clinical doctors. The system distinguishes between domestic and imported cases. The severity of the disease at the time of reporting is also recorded. However, the surveillance system does not follow individual patients over time to record further disease development and final outcome.

Case definition

A case from which *Salmonella* other than *S. Typhi* and *S. Paratyphi* has been isolated or a clinical compatible case with either an epidemiological link to a culture confirmed case or serology indicating recent infection.

Diagnostic/analytical methods used

Bacteriology (isolation of the agent from a clinical sample) followed by confirmation, including serotyping and sometimes genotyping, at the National Reference Laboratory.

Notification system in place

According to the Communicable Disease Act, human cases are notifiable to the Norwegian Surveillance System for Communicable Diseases (MSIS) since 1975.

History of the disease and/or infection in the country

The recorded incidence of salmonellosis in Norway has increased during the last three decades with a sharp rise in the early 1980s due to the emergence of *S. Enteritidis*. In the majority of cases of salmonellosis (approximately 80%), the patients have acquired the disease abroad. The number of reported cases of salmonellosis corresponds well with charter tourism to foreign countries; in years with an increased charter tourism, such as in the mid-1980s and in the period 1992-1998, the incidence of salmonellosis also increased, whereas in years with a lower charter tourism activity due to economical depression, such as in the period 1988-1991, the incidence of salmonellosis dropped.

Since 1998, the incidence of salmonellosis has levelled off. However, an increase was noted during 2001, mostly due to a few large outbreaks. Since 1984, *S. Enteritidis* has become the most common serovar reported, except in 1987 when it was surpassed by *S. Typhimurium* due to a domestic outbreak traced to contaminated chocolate bars. While *S. Typhimurium* predominated in earlier years, *S. Enteritidis* has increased substantially from a low level in 1975-1982 to a higher level from the mid-1990s. No increase of similar magnitude has been observed for any other serovar. The proportion of imported cases of *S. Enteritidis* infections is particularly high (approximately 90% among patients with known place of acquisition) as this pathogen is not established in the Norwegian poultry production. Among domestic cases, *S. Typhimurium* is

the most common serovar. This serovar, although not established among food producing animals in Norway, does occur in the Norwegian environment such as in wild birds and hedgehogs.

Results of the investigation

In 2010, a total of 1367 cases of salmonellosis were reported (incidence rate 27.8 per 100 000), of which 229 (17%) were infected in Norway. Altogether 583 (43%) of the cases were due to *S. Enteritidis*, of which 57 (10%) were infected in Norway. Altogether, 164 (12%) of the cases were due to *S. Typhimurium*, of which 57 (35%) were domestic cases.

National evaluation of the recent situation, the trends and sources of infection

There was a decrease in the overall number of Salmonella-infections in 2010. Most of the reduction is in the number of patients who have contracted the infection abroad.

For domestically acquired infections, 2006 and 2007 were record years when nearly 400 cases contracted salmonellosis inside Norway, - the highest recorded number since 1987. However, both in 2008 and in 2009, there has been a decrease in the number of patients who get the infection without travelling prior to getting ill. This decrease is probably linked to the decrease in cases who contract salmonellosis abroad, since we assume that a number of the domestic cases are secondary cases to imported infections.

In 2010 three small outbreaks of salmonellosis were recorded in Norway. During the autumn, ten persons were reported ill with *Salmonella* Poona and 8 persons with *Salmonella* Napoli. In the same period Sweden also registered several cases of *Salmonella* Poona with identical MLVA profile as the Norwegian cases. An investigation was done in cooperation with Sweden, but the source of infection is so far not found. In addition one small outbreak of *S. Typhimurium* was reported from a private household where meat bought abroad was the suspected source. Domestic outbreaks of salmonellosis recorded in recent years illustrate that many kinds of foods may be involved in outbreaks, also those of non-animal origin, including imported foods.

Relevance as zoonotic disease

The Norwegian Salmonella Control Programmes have documented that live cattle, swine, and poultry in Norway as well as domestically produced food products of animal origin are virtually free from Salmonella. Each year, approximately 75-80% of reported cases of salmonellosis in humans have acquired the infection abroad. This illustrates that domestic food products of animal origin represent a small risk to the consumer in regard to Salmonella, an assumption that is supported by case-control studies.

However, data show that *S. Typhimurium* occurs endemically in the environment representing a risk for spread through wild animals and untreated water. In defined areas, where an endemic situation in the hedgehog and passerine bird populations has been established, annually minor outbreaks and sporadic cases occur.

Additional information

Patients whose work represents a risk for spread of the disease, e.g., in food production and health care, are advised to stay away from work as long as they have symptoms. It is recommended that for these patients three consecutive faecal samples examined after the symptoms have disappeared should be negative before resuming work.

Table Cases of salmonellosis according to place of infection

Serovar	Place of infection				Total
	Norway	Abroad	Unknown	Total	
<i>S. Enteritidis</i>	57	455	71	583	
<i>S. Typhimurium</i>	57	73	34	164	
<i>S Sp Ssp I Gr O:4 Monofasic Variant</i>	30	74	14	118	
<i>S. Newport</i>	6	28	1	35	
<i>S. Stanley</i>	5	26	2	33	
<i>S. Saintpaul</i>	5	19	2	26	
<i>S. Virchow</i>	2	20	3	25	
<i>S. Kentucky</i>	2	17	1	20	
<i>S. Agona</i>	2	15	3	20	
<i>S. Infantis</i>	2	12	4	18	
Others	61	225	39	325	
Total	229	964	174	1367	

Salmonella in foodstuffs

A. *Salmonella* spp. in eggs and egg products

Eggs and egg products are monitored indirectly by monitoring of the layer population, see chapter on *Salmonella* spp. in animals. Additional testing of egg products is carried out by the food business operators as an integral part of their own check procedures.

B. *Salmonella* spp. in poultry meat and products thereof

Poultry meat and products thereof are monitored indirectly by testing all poultry flocks before slaughter see chapter on *Salmonella* in animals. Additional testing at the slaughterhouses or cutting plants is not required. Surveys are performed occasionally.

C. *Salmonella* spp. in red meat (pig, cattle) and products thereof

Monitoring system

At slaughterhouse and cutting plant: The Norwegian *Salmonella* Control Programme: Each year, a number of carcass swabs and lymph node samples are collected randomly from the animal population (pig, cattle) by slaughter and proportional to the slaughterhouses' throughput. The sampling of carcass swabs is described in this chapter, while the sampling of lymph nodes is described in the chapter on *Salmonella* in animals.

Samples of crushed meat are each year collected according to production capacity of cutting plants.

At meat processing plant: Until 1st of March 2010 when the Commission Regulation (EC) No 2073/2005 and Regulation (EC) No 853/2004 of the European Parliament and of the Council came into force, minced meat and meat preparations were sampled according to Council Directive 94/65/EC.

Frequency of the sampling

At slaughterhouse: Detection of an annual prevalence of 0.1% by 95% confidence level.

At cutting plant: According to production capacity: less than 2 tons: twice a year, 2-20tons: once a month, greater than 20 tons: once a week.

At meat processing plant: According to Council Directive 94/65/EC until the new regulations were implemented. Thereafter according to Regulation 2073/2005. The frequencies may, however, vary according to the origin of the raw material and the demonstrated regional or national salmonella prevalence in the corresponding animal populations.

Type of specimen taken

At slaughterhouse: Surface of carcass.

At cutting plant: Crushed meat from equipment or trimmings.

At meat processing plant: Minced meat or meat preparations.

Methods of sampling (description of sampling techniques)

At slaughterhouse: The upper inner part of the hind legs/pelvic entrance and the cut surface area of the abdomen and chest are swabbed, covering an area of approximately 1400 cm² of each carcass.

At cutting plant: Each sample consists of 25 grams of meat.

At meat processing plant: Each sample consists of 25 grams of minced meat or meat preparations.

Definition of positive finding

A positive sample is a sample from which *Salmonella* has been isolated.

Diagnostic/analytical methods used

Bacteriological method: NMKL No 71:1999

Control program/mechanisms/notification system in place

The Norwegian *Salmonella* Control Programme is mandatory. Detection of *Salmonella*, irrespective of serovar, is notifiable.

Measures in case of the positive findings or single cases

Whenever *Salmonella* is detected in samples taken in the National Control Programmes, the competent authorities must be notified without delay. Actions will be taken to identify and eliminate the source of the contamination in order to prevent further spread. However, in the sheep population in some regions, *S. diarizonae* is endemic. When this serovar is detected in sheep, less extensive measures are carried out.

When *Salmonella* is detected in food already on the market, contaminated food will be withdrawn from the market and destroyed or, exceptionally, submitted to processing by a treatment eliminating the hazard. Investigation into the source of the contamination is initiated if relevant. If *Salmonella* is detected in food

controls at the Border Inspection Posts, the consignments will be either rejected or destroyed or, exceptionally, submitted to processing by a treatment eliminating the hazard.

Results of the investigation

Pig meat

In 2010, a total of 1811 carcasses were swabbed, all were negative. None of the samples of crushed meat from pig were positive. For details, see tables.

Cattle meat

In 2010, a total of 1626 carcasses were swabbed, all were negative for *Salmonella*. None of the samples of crushed bovine meat were positive. For details, see tables.

National evaluation of the recent situation, the trends and sources of infection

The Norwegian *Salmonella* Control Programmes document that domestically produced food products of animal origin are virtually free from *Salmonella*. The surveillance data indicate that the overall prevalence is below 0.1%.

Relevance of the findings in animals to findings in foodstuffs and to human cases

Red and white meat produced in Norway is virtually free from *Salmonella*, and the risk of contracting *Salmonella* from domestically produced animal products is small. A connection between meat or meat products of domestic origin and human infection has never been established.

Table *Salmonella* in foodstuffs

	Source of information	Sample weight	Units tested	Total units positive for <i>Salmonella</i> spp.
Pig - carcass swabs	NSCP ¹	swab	1811	0
Cattle - carcass swabs	NSCP	swab	1626	0
Crushed red meat (pig, cattle)	NSCP	25g	1656	0
Fish - wild catch	NIFES	25g	172	0
Fish - farmed	NIFES	25g	20	0
Fish oil	NIFES	25 g	17	0
Crustaceans - raw	NIFES	25g	15	0
Crustaceans - cooked	NIFES	25g	14	0
Molluscan shellfish - raw	NIFES	25g	99	0
Molluscan shellfish - cooked	NIFES	25g	1	0

¹NSCP = Norwegian *Salmonella* Control Programme

Salmonella in animals

A. *Salmonella* spp. in poultry (*Gallus gallus*, turkeys, ducks, geese, guinea fowls)

Monitoring system

Breeding flocks

The Norwegian *Salmonella* Control Programme established pursuant to Article 5 of Regulation (EC) 2160/2003 and approved by the EFTA Surveillance Authority (ESA) (364/07/COL) includes all poultry breeding flocks. Sampling takes place at the initiative of the food business operator and by the Competent Authority according to Regulation (EC) 1003/2005 (in force throughout 2010). The Norwegian *Salmonella* Control Programmes also include all breeder flocks of ducks, geese, turkeys and guinea fowl. Other strategies: Animals are tested in relation to clinical surveillance and import. Norway is also granted additional guarantees according to Commission decision 2003/644/EC.

Laying hens flocks

The Norwegian *Salmonella* Control Programme: All laying hen flocks are tested at the farm. Sampling takes place at the initiative of the food business operator and by the Competent Authority according to Regulation (EC) 1168/2006.

Other strategies: Animals are tested in relation to clinical surveillance and import. Additional guarantees according to Commission decision 2004/235/EC also applies to Norway.

Meat producing flocks (broilers, turkeys, ducks, geese, guinea fowl)

The Norwegian *Salmonella* Control Programme: All poultry flocks are tested before slaughter. Sampling takes place at the initiative of the food business operator and one a year by the Competent Authority according to Regulation (EC) 646/2007. If poultry for slaughter are imported, additional guarantees according to 95/410/EC applies.

Frequency of the sampling

Breeding flocks

Day-old chicks: Every flock is sampled.

Rearing period: Every flock is sampled twice.

Production period: Every second week.

Laying hens

Day-old chicks: Every flock is sampled.

Rearing period: 2 weeks prior to moving.

Production period: Every 15 weeks.

Before slaughter at farm: Every flock for slaughter is sampled.

Meat producing flocks (broilers, turkeys, ducks, geese, guinea fowl)

Before slaughter at farm: Every flock is sampled

Type of specimen taken

Breeding flocks

Day-old chicks: Internal linings of delivery boxes.

Rearing period: Socks/boot swabs.

Production period: Socks/boot swabs or faeces (caged birds) and dust swabs.

Before slaughter at farm: Socks/boot swabs or faeces (caged birds) and dust swabs.

Laying hens

Day-old chicks: Internal linings of delivery boxes.

Rearing period, production period: Socks/boot swabs or faeces (caged birds).

Before slaughter at farm: Socks/boot swabs or faeces (caged birds).

Meat producing flocks (broilers, turkeys, ducks, geese, guinea fowls)

Before slaughter at farm: Socks/boot swabs and dust swabs.

Methods of sampling (description of sampling techniques)

Breeding flocks

Day-old chicks: All flocks: Transport crates are tested (crate liners or swabs).

Rearing period: All flocks: Tested at 4 weeks of age and 2 weeks before moving by two pairs of socks.

Production period: All flocks: Tested every 2nd week by two pairs of socks (caged birds: faecal samples) and one dust sample.

Laying hens

Day-old chicks: All flocks: Transport crates are tested (crate liners or swabs).

Rearing period: All flocks: Tested two weeks before moving by 2 pair of socks (caged birds: faeces).

Production period: All flocks: Tested every 15 weeks by two pairs of socks (caged birds: faeces).

Before slaughter: All flocks for slaughter: Tested before slaughter by 2 pair of socks (caged birds: faeces).

Meat producing flocks (broilers, turkeys, ducks, geese, guinea fowls)

Before slaughter at farm: Every flock is sampled by one pair of socks and one dust sample.

Case definition

A positive flock is a flock from which *Salmonella* (irrespective of serovar) has been isolated from at least one sample.

Diagnostic/analytical methods used

Bacteriological method: ISO 6579:2002.

Vaccination policy

Vaccination against *Salmonella* is prohibited in Norway.

Control program/mechanisms/notification system in place

The Norwegian *Salmonella* Control Programme is mandatory. Detection of *Salmonella*, irrespective of serovar, has been notifiable since 1965.

Measures in case of the positive findings or single cases

Whenever *Salmonella* is detected, the competent authorities must be notified without delay. Also, relevant food business operators, such as slaughterhouses, hatcheries, and egg collecting centres receiving animals or animal products from an infected animal holding must be informed. Stringent restrictions including cleaning and disinfection, control of animal movement and control of person admission will be imposed on an infected animal holding. Infected animals must be isolated from other animals. Whenever *Salmonella* is detected, epidemiological investigations also including the feed suppliers will be initiated in order to identify and eliminate the source of infection. If *Salmonella* is detected, the whole flock will be destroyed or subjected to sanitation slaughter. Eggs from hatcheries where *Salmonella* has been detected will be destroyed or pasteurized. If *Salmonella* is detected in chicks, all chicks from the same hatchery machine must be destroyed. Farms that have received infected chicks will be considered infected and restrictions will be imposed on these farms as well. Restrictions will be lifted when infected rooms have been cleaned and disinfected, bacteriological testing following cleaning and disinfection gives a negative test result, and the rooms have been empty for at least 30 days.

Results of the investigation

Gallus gallus breeding flocks

In 2010, a total of 105 rearing flocks and 156 production flocks were tested, all were negative for *Salmonella*. For details, see table.

Gallus gallus laying hen flocks

In 2010, a total of 146 rearing and 981 adult flocks were tested, all were negative. For details, see table.

Gallus gallus broiler flocks

In 2010, 4549 broiler flocks (batches for slaughter) were investigated, and two were positive (*S. Senftenberg* and *S. Brandenburg* respectively). These two flocks were also positive in the follow up sampling. For details, see table.

Other poultry (turkeys, ducks, geese, guinea fowls)

In 2010, none of the Norwegian breeder flocks were positive. None of the production flocks were positive.

In addition to the Control Programme, samples have been taken in relation to clinical problems, follow up or various projects. None of these samples were positive for *Salmonella*. For details, see table.

National evaluation of the recent situation, the trends and sources of infection

The favourable *Salmonella* situation in Norwegian poultry is partly dependent upon an efficient control of breeding flocks. Due to extensive surveillance during many years, stringent measures in case of positive findings, and restricted import, poultry breeding flocks in Norway are virtually free from *Salmonella*. *S. Agona* was found in a broiler parent flock in 2001. *S. Enteritidis* was for the first time detected in Norwegian poultry production in a broiler flock in 2007. *S. Enteritidis* has never been detected in Norwegian breeding flocks or in laying hens. The duck, geese, turkey and guinea fowl populations in Norway are small. A

few times, positive commercial flocks have been found, the last time in 2000, when two turkey flocks were positive for *S. Aberdeen* and *S. Typhimurium*, respectively.

Relevance of the findings in animals to findings in foodstuffs and to human cases

The Norwegian Salmonella Control Programmes have documented that so far poultry in Norway as well as domestically produced poultry products are virtually free from *Salmonella*. Each year, approximately 75-80% of reported cases of salmonellosis in humans have acquired the infection abroad. This illustrates that domestic food products of animal origin represent a small risk to the consumer in regard to *Salmonella*, an assumption that is supported by case-control studies.

Table *Salmonella* in poultry and other birds

	Source of information	Sampling unit	Total units tested (official and industry sampling)	Total units positive for <i>Salmonella</i> spp.
Gallus gallus (fowl)				
Grandparents (egg line) – production period	NSCP ¹	flock	3	0
Parents - egg line – rearing period	NSCP	flock	14	0
Parents - egg line – production period	NSCP	flock	20	0
Parents - meat line – rearing period	NSCP	flock	91	0
Parents - meat line – production period	NSCP	flock	136	0
Laying hens – rearing period	NSCP	flock	146	0
Laying hens – production period	NSCP	flock	981	0
Broilers	NSCP	batch	4549	2 ²
Unspecified	NVI	holding	28	2 ³
Ducks				
Breeding flocks	NSCP	flock	4	0
Meat production flocks	NSCP	batch	74	0
Unspecified	NVI	flock	4	0
Geese				
Meat production flocks	NSCP	batch	3	0
Unspecified	NVI	flock	1	0
Turkeys				
Breeding flocks	NSCP	flock	15	0
Meat production flocks	NSCP	batch	385	0
Unspecified	NVI	flock	12	0
Quails	NVI	holding	2	1 ⁴
Other domestic birds (mainly pet psittacine birds)	NVI	animal	21	0
Wild birds	NVI	animal	14	2 ⁵

¹ NSCP = Norwegian *Salmonella* Control Programme

² *S. Senftenberg* (1), *S. Brandenburg* (1).

³ *S. Senftenberg* (1), *S. Brandenburg* (1). Both found after repeated sampling in holdings/flocks found positive in NSCP.

⁴ *S. Infantis*.

⁵ *S. Typhimurium* (2).

Footnote: The samples reported from NVI (Norwegian Veterinary Institute) are mainly from clinical investigations, but might also represent import controls and other reasons for sampling.

B. *Salmonella* spp. in pigs and cattle

Monitoring system

Pig breeding herds: The Norwegian *Salmonella* Control Programme: All elite breeding herds are tested.

Pig multiplying herds: The Norwegian *Salmonella* Control Programme: Each year, a number of lymph node samples and carcass swabs are collected randomly from the sow population at slaughterhouse according to the slaughter volume. The sampling of lymph nodes is described in this chapter, the sampling of carcass swabs is described in the chapter on *Salmonella* in foodstuffs.

Cattle and pig fattening herds: The Norwegian *Salmonella* Control Programme: Each year, a number of lymph node samples and carcass swabs are collected by slaughter and proportionally distributed according to the slaughterhouses' capacities. The sampling of lymph nodes is described in this chapter, while the sampling of carcass swabs is described in the chapter on *Salmonella* in foodstuffs.

Other strategies: Animals are tested in relation to clinical surveillance and import.

Frequency of the sampling

Pig breeding herds: Once a year.

Animals at slaughter (herd based approach): Detection of an animal prevalence level of 0.1% by 95% confidence.

Type of specimen taken

Pig breeding herds: Faeces.

Animals at slaughter (herd based approach): Lymph nodes.

Methods of sampling (description of sampling techniques)

Animals at farm: If there are clinical problems with diarrhoea, faecal samples will be taken.

Pig breeding herds: Faecal samples.

Animals at slaughter (herd based approach): From each carcass at least five ileocaecal lymph nodes are aseptically removed and pooled in a plastic bag. All samples are kept refrigerated during the period of sampling and sent to the laboratory the same day.

Case definition

A positive sample is a sample from which *Salmonella* has been isolated.

Diagnostic/analytical methods used

Faeces: Annex D of ISO 6579.

Lymph nodes: NMKL No 71:1999

Vaccination policy

Vaccination against *Salmonella* is prohibited in Norway.

Control program/mechanisms/notification system in place

The Norwegian *Salmonella* Control Programme is mandatory. Detection of *Salmonella*, irrespective of serovar, has been notifiable since 1965.

Measures in case of the positive findings or single cases

Whenever *Salmonella* is detected, the competent authorities must be notified without delay. Actions will be taken to identify and eliminate the source of the contamination in order to prevent further spread. Also, slaughterhouses and food production facilities receiving animals or animal products from an infected animal holding must be informed. Stringent restrictions including cleaning and disinfection, control of animal movement and control of person admission will be imposed on an infected animal holding. Infected animals must be isolated from other animals. Animals are not allowed to be sent to slaughter without permission from the Food Safety Authority and if sent to slaughter, the slaughterhouse must be notified so that sanitation slaughtering can be conducted. Milk from infected cattle herds must be pasteurised. Whenever *Salmonella* is detected, epidemiological investigations also including the feed suppliers will be initiated in order to identify and eliminate the source of infection. There will be intensified sampling, also on farms that have had contact with the infected holding. Restrictions will be lifted when all animals have been tested with a negative test result in two consecutive samplings with a minimum interval of 30 days. Following lifting of the restrictions, retesting will be conducted after approx. six months.

Results of the investigation

Pigs

In 2010, lymph node samples from a total of 2226 animals were tested in the Norwegian Salmonella Control Programme. One sample was positive for *S. Bovismorbificans*. None of the 117 herds tested with faecal samples (2090 samples in total) were positive.

In addition, 628 samples from 61 different herds were investigated, mainly due to follow up of positive findings. Three herds were positive (*S. Typhimurium* (2), *S. Bovismorbificans* (1)). The herd positive for *S. Bovismorbificans* was the same as the herd found positive for *S. Bovismorbificans* in the Norwegian Salmonella Control Programme. This herd also had cattle positive for *S. Bovismorbificans*. The two herds with *S. Typhimurium* had contact with each other.

Cattle

In 2010, a total of 1854 animals were sampled (lymph node samples) in the Norwegian Salmonella Control Programme. All samples were negative for *Salmonella*.

In addition, 1266 samples from 186 different herds were investigated, mainly due to follow up of positive findings. Three herds were positive for *S. Bovismorbificans*. One of the herds also had pigs positive for *S. Bovismorbificans*.

National evaluation of the recent situation, the trends and sources of infection

The Norwegian *Salmonella* Control Programmes document that Norwegian food producing animals are virtually free from *Salmonella*. The surveillance data indicate that the overall prevalence is below 0.1%.

C. *Salmonella* spp. in other animals

Monitoring system

Described here is *Salmonella* in sheep and goats and other animal species than food producing animals, such as pets, zoo animals, reptiles and wild life. Sampling is done in relation to clinical surveillance and import.

Case definition

A positive animal is an animal from which *Salmonella*, irrespective of serovar, has been isolated.

Vaccination policy

Vaccination against *Salmonella* is prohibited in Norway.

Measures in case of the positive findings or single cases

Whenever *Salmonella* is detected, the competent authorities must be notified without delay. Unless the finding is in a wild animal, epidemiological investigations will be initiated in order to identify and eliminate the source of infection.

Notification system in place

Detection of *Salmonella*, irrespective of serovar, has been notifiable since 1965.

Results of the investigation

For details see table. In addition to the results presented above and in the tables, animals may have been sampled due to clinical problems, follow up or various projects. None of these samples have been positive.

Relevance of the findings in animals to findings in foodstuffs and to human cases

A substantial proportion of the *S. Typhimurium* infections in humans are indigenous. This serovar, although not established among food animals in Norway, does occur in Norwegian wild birds and hedgehogs, and these two sources have been described to be the source for almost half of all indigenous *S. Typhimurium* cases. These two sources probably also constitutes a risk for food producing animals. Also, reptiles kept as pets pose a risk for transmission to humans.

Table *Salmonella* in animals other than birds

	Source of information	Sampling unit	Units tested	Total units positive for <i>Salmonella</i> spp.	<i>S. Typhimurium</i>	<i>S. Dublin</i>	<i>S. Bovismorbificans</i>	<i>S. diarizonae</i> (61:k:1,5,(7))	<i>Salmonella</i> spp.
Cattle									
At slaughterhouse, lymph nodes	NSCP ¹	animal	1854	0					
Other investigations ²	NVI	herd	186	3			3		
Sheep									
Clinical investigations ³	NVI	animal	103	18				18	
Pigs									
Breeding animals – at farm	NSCP	herd	117	0					
Slaughtered animals – lymph nodes	NSCP	animal	2226	1			1		
Other investigations ⁴	NVI	herd	61	3	2		1		
Solipeds, domestic	NVI	animal	54	0					
Alpacas	NVI	animal	67	0					
Cats ⁵	NVI	animal	126	2		2			
Dogs ⁶	NVI	animal	453	11	2				9
Pets (rabbit, chinchilla etc.)	NVI	animal	14	0					
Reptiles and turtles ⁷	NVI	animal	8	5					5
Wild animals ⁸	NVI	animal	4	1					1
Zoo animals ⁹	NVI	animal	38	6					6

¹ NSCP = Norwegian *Salmonella* Control Programme.

² A total of 1266 samples.

³ A total of 50 sampled herds.

⁴ A total of 628 samples.

⁵ The two positive animals were two aborted fetuses from the same household.

⁶ The 9 positive dogs reported in column *Salmonella* spp. were positive for: *S. Brandenburg* (1), *S. enterica* subsp. *enterica* 4,5,12:i:- (3), *S. Infantis* (1), *S. Livingstone* (1), *S. Putten* (1), *S. Meleagridis* (1), double infection with *S. Brandenburg* and *S. Enteritidis* (1).

⁷ Excluding reptiles from zoos. The 5 positive animals reported in column *Salmonella* spp. were positive for: *S. Muenchen* (2), *S. Paratyphi B* (1), *S. enterica* subsp. *indica* (1), triple infection with *S. Tennessee* + *S. Cotham* + *S. arizonae* 44:z4z32:- (1).

⁸ The serovar reported in column *Salmonella* spp. is: *S. enterica* subsp. *diarizonae* 38 : k : z35 found in a Raccoon Dog.

⁹ Animals from 8 zoos and similar holdings. The serovars reported in column *Salmonella* spp. are: *S. Nyeko* (1), *S. Senftenberg* (1), *S. Muenchen* (1), *S. Wedding* (1), double infection with *S. Schwarzengrund* and *S. Yoff* (1), *S. enterica* subsp. *houtenae* (1).

Footnote: The samples reported from NVI (Norwegian Veterinary Institute) include clinical investigations, follow up of positive findings, import controls and other reasons for sampling. Animal species where the number of samples were <10 and all samples were negative are not reported.

Salmonella in feedingstuffs

History of the disease and/or infection in the country

Norway has for many years performed an extensive surveillance of feedingstuffs and imposed stringent measures in case of positive findings. The import of animal feedingstuffs has also been restricted for many years. The result is that the feedingstuffs given to Norwegian livestock for many years have virtually been free from *Salmonella*.

National evaluation of the recent situation, the trends and sources of infection

Extensive surveillance systems for *Salmonella* in regard to feedingstuffs are established in order to prevent animals from being exposed to contaminated feed. Feedingstuffs for both terrestrial animals and fish are covered by surveillance programmes. The surveillance programmes document a low prevalence of *Salmonella* in domestically produced animal compound feedingstuffs. However, data from process control, including environmental sampling, indicates that there are certain serovars that sometimes contaminate production facilities, especially those producing fish feed.

Relevance of the findings in animals, feedingstuffs and foodstuffs to human cases

The favourable *Salmonella* situation in animals and humans in Norway is partly dependent upon the efficient control of animal feedingstuffs. The number of animals infected through feedingstuffs is probably very low, and this route of infection probably represents a negligible risk to humans.

Recent actions taken to control the zoonoses

Detection of *Salmonella* is notifiable and the establishment must take immediate actions to prevent the distribution of contaminated feed. Contaminated feed will either be destroyed, heat or acid treated.

In general, complete feedingstuffs and protein concentrates (supplementary feedingstuffs) intended for poultry, pigs, and cattle are exposed to heat treatment of at least 81 degrees Celsius core temperature and the production has to take place in a production line where all the other feedingstuffs are heat treated. According to the regulations for production of feedingstuffs, feed mills are required to have in place a system to monitor the acceptability of the process including a sampling scheme for *Salmonella* consisting of minimum 3 samples every fortnight (all poultry feed mills and pig and cattle feed mills with a capacity above 10,000 tons per year) or every fourth week (pig and cattle feed mills with a capacity below 10,000 tons per year). Samples include raw materials and scrapings from control points. Establishments preparing feed for fur animals are required to analyse a minimum of one sample for *Salmonella* per month. The national production of meat and bone meal is subject to a continuous process control that includes analyses for *Salmonella*. Through an official surveillance programme random samples of feedingstuffs for terrestrial animals are collected and analysed for the presence of *Salmonella*.

Establishments producing fish feed are required to establish and maintain an internal (process) control based on the HACCP-system according to the regulation for fish feed. A minimum of four samples per 14 days should be examined with respect to *Salmonella*. Through an official surveillance programme described in the regulation for feedingstuffs for fish, random samples of feedingstuffs for fish are collected at the establishments and analysed for the presence of *Salmonella*.

Establishments producing fish meal or fish oil are required to establish and maintain an internal (process) control based on the HACCP-system according to the regulation for fish meal and fish oil. A minimum of one sample per 50 tons must be tested for the presence of *Salmonella*.

Imported feed materials of vegetable origin must be subjected to control for *Salmonella* before distribution or use. The number of samples depends on the size of the load and whether the feedingstuffs are classified as high-risk (soy beans, maize, cotton seed, etc.) or low-risk materials. Imported feed of animal origin, predominantly pet feed, is controlled at one of the Border Inspection Posts according to Council Directives 97/78/EEC and 89/662/EEC. Dog treats made from hides that are imported from third countries must be accompanied with a certificate that documents that the lot has been controlled for *Salmonella*. At the Border Inspection Posts, sampling is done according to a specific scheme. Feed materials, including fish meal, imported from third countries must be subjected to control for *Salmonella* according to a specified plan before distribution or use. A minimum of one sample per 50 tons must be tested for the presence of *Salmonella*.

In addition to the surveillance run by the government or the industry itself, feedingstuffs are also subjected to analyses for *Salmonella* in relation to epidemiological investigations and specific surveys and studies.

Table *Salmonella* in feedingstuffs

	Source of information	Total units tested	Units tested in official controls	Units tested in surveillance by industry	Total units positive for <i>Salmonella</i> spp. ¹
Feed matter					
Barley derived	NFSA	24	0	24	0
Oat derived	NFSA	11	0	11	0
Wheat derived	NFSA	461	0	461	0
Maize (including maize derived)	NFSA	336	0	336	0
Rape seed derived	NFSA	268	0	268	1
Soya(bean) derived	NFSA	3136	0	3136	59
Sunflower seed derived	NFSA	36	0	36	0
Legume seeds and similar products	NFSA	54	0	54	0
Tubers, roots, other seeds	NFSA	12	0	12	0
Other feed material	NFSA	2	2	0	0
Meat and bone meal	NFSA	277	0	277	4
Fish meal	NFSA	96	0	96	3
Fish oil	NFSA	1	0	1	0
Compound feedingstuffs for					
swine	NFSA	34	34	0	0
poultry	NFSA	44	44	0	0
cattle, swine, poultry	NFSA	457	0	457	0
fish	NFSA	1798	0	1798	191
fur animals	NFSA	381	0	381	0
horses	NFSA	1	1	0	0
Environmental samples including process control samples – not specified to production	NFSA	13385	149	13236	146

¹ The positive samples were mainly imported feed material or environmental samples.

Antimicrobial resistance in *Salmonella* isolates

Sampling strategy used in monitoring

All *Salmonella* found in production animals, irrespective if they are found in the Norwegian *Salmonella* Control Programmes or in connection with clinical problems, surveys or other investigations, are included in the resistance monitoring (only one isolate per herd). *Salmonella* isolated from other animals may be susceptibility tested as well. Exceptions from the rules described above are that not all *S. diarizonae* from sheep or *S. Typhimurium* from wild birds and wild animals or *Salmonella* from reptiles, wild animals or zoo animals are tested every year.

Type of specimen taken

Salmonella isolates collected through the Norwegian *Salmonella* Control programmes, which include those animal species required by the Commission Decision No 2007/407/EC on the harmonised monitoring of antimicrobial resistance in *Salmonella*. Isolates from other samples taken vary depending on the situation.

Methods of sampling (description of sampling techniques)

For description of the Norwegian *Salmonella* Control programmes, see the parts describing *Salmonella* in the various animal species. Other sampling methods vary depending on the situation.

Procedures for the selection of isolates for antimicrobial testing and methods used for collecting data

One isolate per herd is selected for antimicrobial testing. *Salmonella* is isolated at various laboratories and sent to the Norwegian Veterinary Institute in Oslo for testing of antimicrobial susceptibility.

Laboratory methodology used for identification of the microbial isolates

Normally, ISO 6579:2002 or NMKL No 71:1999 are used for isolation of *Salmonella*. However, isolates may have been obtained by other methods as well.

Laboratory methods used for detection for resistance

The VetMIC microdilution method (Dept. of Antibiotics, National Veterinary Institute, Sweden) is used for the susceptibility testing of all isolates. The antimicrobials included are listed in the tables.

Breakpoints used in testing

For interpretation of results epidemiological cut-off values recommended by EFSA were applied.

Control program/mechanisms

The resistance testing of *Salmonella* isolated from animals is a part of the Norwegian monitoring programme for antimicrobial resistance in feed, food and animals - NORM-VET.

Results

The qualitative data are presented in the table. Quantitative data as well as data on breakpoints and range of testing are presented in the NORM/NORM-VET 2010 report.

Table Antimicrobial susceptibility testing of *Salmonella* spp.

	S. Bovismorbificans – cattle and swine	S. Typhimurium – swine	S. Senftenberg – broiler	S. Brandenburg – broiler
Number of isolates tested	4	2	1	1
Number of isolates resistant to				
Tetracycline	0	1	0	0
Chloramphenicol	0	0	0	0
Cefotaxime	0	0	0	0
Ciprofloxacin	0	0	0	0
Nalidixic acid	0	0	0	0
Sulfamethoxazole	0	0	0	0
Trimethoprim	0	2	0	0
Streptomycin	0	2	0	0
Gentamicin	0	0	0	0
Ampicillin	0	1	0	0

Campylobacteriosis

General evaluation of the national situation

History of the disease and/or infection in the country

Norwegian studies have shown that many species of wild birds, especially crows and seagulls, are frequent carriers of thermophilic *Campylobacter* spp. Thermophilic *Campylobacter* spp. have also been isolated from poultry, dogs, cats, pigs, sheep, cattle, and flies, and sporadically from wild mammals. Before 2001, when the surveillance programme in broilers was implemented, the prevalence of thermophilic *Campylobacter* spp. in Norwegian broiler flocks had been studied twice. In 1990, 18% of the flocks tested were infected, whereas this proportion in 1997-1998 had decreased to 4%. This reduction was attributed to an increased focus on the importance of biosecurity.

The Action Plan against *Campylobacter* in broilers that started in 2001 has shown that the yearly incidence of broiler flocks being positive for *Campylobacter* has varied between 3.3% and 6.3% in the years 2002-2007. The data from 2008 - 2010 are not directly comparable to previous years because in 2008 the sampling was reduced to sampling prior to slaughter only and in 2009 the surveillance was altered from full year surveillance to the period between 1st May to the end of October when the incidence is highest. estimated full year prevalence of positive flocks in 2008 was probably approximately the same as in 2007 and the results from 2009 and 2010 probably gradually improving.

The number of flocks going positive out on the market has been reduced from 127 in 2002 to 58 in 2007. The number of positive flocks out on the market was probably similar to 2007 in 2008 and slightly higher in 2009 and 2010.

In 1998, campylobacteriosis for the first time surpassed salmonellosis as the most frequently reported bacterial cause of acute human gastroenteritis in Norway, and since then the reported incidence of campylobacteriosis has been above that of salmonellosis. Since the beginning of the 1990s and until it peaked in 2001, there was a major increase in the incidence of campylobacteriosis in Norway, both in domestic and imported cases. Usually, 50-60% of the cases are imported.

National evaluation of the recent situation, the trends and sources of infection

The reported human incidence in 2010 was slightly lower than the incidence reported in 2009. The data on prevalence in broiler flocks in 2010 were not as complete as the data from the period 2001 - 2007, but we assume that there is no major change in the prevalence. We also assume that in 2010, as in earlier years, the majority of the positive flocks were detected before slaughter, and were therefore treated (i.e. frozen or heat treated) before they went on the market. The use of untreated water is considered an important source of campylobacteriosis in Norway.

Relevance of the findings in animals, feedingstuffs and foodstuffs to human cases

The poultry production and poultry consumption has increased during the last years. Even if the Norwegian action plan against *Campylobacter* in broilers have largely reduced the number of *Campylobacter* positive broiler carcasses entering the market, there are still positive broiler carcasses on the market. In addition, other food products may also be positive for *Campylobacter*. An important source of human campylobacteriosis in Norway is the use of untreated water, in private homes and cottages and during camping and hiking.

Recent actions taken to control the zoonoses

The implementation of the Norwegian action plan against *Campylobacter* in broilers in 2001 was a direct response from the authorities, scientific institutions and the industry to the major increase in human campylobacteriosis that was seen during the late 1990s and up to 2001.

Campylobacteriosis in humans

Reporting system in place for the human cases

Human cases are reported to the Norwegian Surveillance System for Communicable Diseases (MSIS), from microbiological laboratories as well as from clinical doctors. The system distinguishes between domestic and imported cases. The severity of the disease at the time of reporting is also recorded. However, the surveillance system does not follow individual patients over time to record further disease development and final outcome.

Case definition

A case from which *Campylobacter* spp. has been isolated or a clinical compatible case with an epidemiological link to a culture confirmed case.

Diagnostic/analytical methods used

Bacteriology (isolation of *Campylobacter* species from faecal samples) followed by voluntary confirmation (species identification and biotyping) at the National Reference Laboratory. Due to the methods applied, *C. lari* and *C. upsaliensis* are probably underdiagnosed.

Notification system in place

According to the Communicable Disease Act, human cases are notifiable to the Norwegian Surveillance System for Communicable Diseases (MSIS) since 1991.

History of the disease and/or infection in the country

Since the beginning of the 1990s and until it peaked in 2001, there was a significant increase in the incidence of campylobacteriosis in Norway. From 1997 to 2001, the incidence increased by ~145%. In 1998, campylobacteriosis for the first time surpassed salmonellosis as the most frequently reported bacterial cause of acute gastroenteritis in Norway, and since then the reported incidence of campylobacteriosis has been above that of salmonellosis. Usually, 50-60% of the cases are imported. The increased incidences observed throughout the 1990s and until 2001 were due to a rising number of both domestic and imported cases. The number of cases, both domestic and imported declined in 2002 and was stable during the period from 2002 to 2004. In 2005, the number of cases increased again and the number of domestic and imported cases were for the first time almost the same. In 2006 the number of imported cases were stable and the number of domestic cases decreased compared to 2005. Also in 2007, the incidence of imported cases raised compared to the domestic cases. The number of imported cases decreased again in 2009 and the number of domestic cases increased to the same level as in 2001 and 2005. Most cases are sporadic. A case control study conducted in Norway during 1999-2000 identified consumption of untreated drinking water, consumption of poultry meat purchased fresh, consumption of barbecued meat, and professional contact with animals as significant risk factors in regard to campylobacteriosis. Daily contact with dogs/cats was identified as a risk factor in case control studies conducted during the early 1990s, but was not identified as a risk factor in the 1999-2000 study. Studies indicate that the vast majority (~95%) of reported cases are due to *C. jejuni*, and that *C. coli* is the cause of most of the remaining cases.

Results of the investigation

In 2010, a total of 2673 cases (incidence rate 54.3 per 100 000) were reported of which 1371 (51%) were known to be imported, 998 (37%) were domestic and 304 (11%) had an unknown place of infection. Altogether, five smaller foodborne outbreaks of campylobacteriosis were registered. No deaths due to campylobacteriosis were reported.

National evaluation of the recent situation, the trends and sources of infection

The number of reported domestic cases decreased in 2010 compared to 2009. The incidence of domestic human campylobacteriosis has been relatively stable during the last five years. As the overall occurrence of positive broiler flocks is low, there must be other important sources to human campylobacteriosis apart from poultry products in Norway, untreated drinking water probably being the most important one.

Relevance as zoonotic disease

Campylobacter is the most frequently reported cause of bacterial gastroenteritis in Norway. Every year, approximately half of the reported cases have acquired the infection in Norway.

Additional information

Patients whose work represents a risk for spread of the disease, e.g., in food production and health care, are advised to stay away from such work while they are having symptoms. It is recommended that for these patients two consecutive faecal samples examined after the symptoms have disappeared should be negative before returning to work.

Table Campylobacteriosis cases infected in Norway 2006 - 2010. Incidence rate (IR) per 100.000 inhabitants 2010.

County	2006	2007	2008	2009	2010	IR 2010
Østfold	31	37	62	52	34	12.4
Akershus	90	92	99	88	89	16.3
Oslo	80	93	86	129	108	18.0
Hedmark	42	31	43	53	38	19.8
Oppland	57	39	62	45	31	16.7
Buskerud	58	47	47	65	42	16.1
Vestfold	43	36	69	47	57	24.4
Telemark	32	36	33	41	25	14.8
Aust-Agder	33	16	20	13	9	8.2
Vest-Agder	32	27	20	22	27	15.7
Rogaland	170	198	157	162	145	33.3
Hordaland	159	154	125	174	127	26.2
Sogn & Fjordane	28	26	20	44	24	22.3
Møre & Romsdal	50	53	39	64	55	21.7
Sør-Trøndelag	87	114	107	120	92	31.3
Nord-Trøndelag	20	39	36	40	39	29.5
Nordland	33	50	33	40	30	12.6
Troms	42	50	33	34	14	8.9
Finnmark	10	15	11	22	12	16.3
Total	1097	1153	1102	1255	998	20.3

Campylobacter in foodstuffs

Thermophilic *Campylobacter* in broiler meat and products thereof

Monitoring system

See chapter on *Campylobacter* in *Gallus gallus*.

Methods of sampling/definition of positive finding

See chapter on *Campylobacter* in *Gallus gallus*.

Preventive measures in place

In the surveillance programme, the broiler flocks found positive before slaughter are subjected to freezing for at least 3 weeks or heat treatment.

Control program/mechanisms

The Norwegian action plan against *Campylobacter* in broilers is a surveillance programme agreed upon by the Norwegian Food Safety Authority, scientific institutions and the poultry industry.

Recent actions taken to control the zoonoses

The establishment of the Norwegian action plan against *Campylobacter* in broilers was a direct response to the major increase in the incidence of human campylobacteriosis during the 1990s.

Measures in case of the positive findings or single cases

See chapter on *Campylobacter* in *Gallus gallus*.

Notification system in place

All findings in the Norwegian action plan against *Campylobacter* in broilers are reported and published as summary reports.

Results of the investigation

The results from the Norwegian action plan against *Campylobacter* in broilers are presented in the chapter on *Campylobacter* in *Gallus gallus*.

National evaluation of the recent situation, the trends and sources of infection

The Norwegian campylobacteriosis situation is a concern for the authorities. The establishment of the Norwegian action plan against *Campylobacter* sp. in broilers in 2001 was a response to the urgent situation. This action plan has since it was established and through 2010 prevented millions of *Campylobacter* positive broiler carcasses from entering the market raw.

Campylobacter in animals

Thermophilic *Campylobacter* in *Gallus gallus*

Monitoring system

A surveillance programme in broilers was implemented in May 2001 (part of the Norwegian action plan against *Campylobacter* in broilers).

Frequency of the sampling

Before slaughter at farm: Between 1 May and 31 October, which corresponds with the high season for *Campylobacter* positive flocks, every flock is sampled.

At slaughter: Flocks where the result from the pre slaughter sample is lacking at the time of slaughter are sampled by staff at the Norwegian Food Safety Authority.

Type of specimen taken

Before slaughter at farm: Faeces

At slaughter: Caecum

Methods of sampling (description of sampling techniques)

Before slaughter at farm: 10 swabs from fresh faecal droppings are taken by the owner maximum four days before slaughter. They are transported dry as one pooled sample to the laboratory.

At slaughter: 10 caecae are sampled at the slaughter line. The 10 samples are pooled to one at the laboratory.

Case definition

Before slaughter at farm: A flock where *Campylobacter* spp. is found.

At slaughter: A slaughter batch where *Campylobacter* spp. is found.

Diagnostic/analytical methods used

Before slaughter at farm: PCR Real Time PCR

At slaughter: ISO 10272-1:2006.

Vaccination policy

There is no vaccination against *Campylobacter* in Norway.

Other preventive measures than vaccination in place

Farms producing *Campylobacter* positive flocks are subject to follow-up visits from the advisors in the industry and veterinary supervisors from the Norwegian Food Safety Authority to assist in implementing measures preventing further flocks to be infected with *Campylobacter*.

Control program/mechanisms

The Norwegian action plan against *Campylobacter* in broilers is a surveillance programme agreed upon by the Norwegian Food Safety Authority, scientific institutions and the poultry industry. The surveillance programme is compulsory.

Recent actions taken to control the zoonoses

The establishment of the Norwegian action plan against *Campylobacter* in broilers was a direct response to the major increase in the incidence of human campylobacteriosis during the 1990s.

Measures in case of the positive findings or single cases

Carcasses from flocks that are positive for thermophilic *Campylobacter* sp. based upon the pre-slaughter sampling are either subjected to heat-treatment or frozen for a minimum of three weeks. The poultry industry uses data from the surveillance programme as an incentive for improving the hygienic conditions on broiler farms.

Notification system in place

All positive flocks in the surveillance programme are reported to the authorities.

Results of the investigation

In 2010, in the period 1 May - 31 October, a total of 2170 samples (representing approx 2170 flocks, and covering virtually all slaughtered flocks in Norway in that period) were taken approximately four days before slaughter. A total of 110 samples (5.1%) were positive for *Campylobacter* spp. In addition - one sample (negative) was taken at slaughter due to lack of results from the pre-slaughter sample.

National evaluation of the recent situation, the trends and sources of infection

The poultry production has increased in Norway during the last years. The yearly prevalence of flocks being positive for *Campylobacter* from 2002 to 2007 was between 3.3 and 6.3%. The results from 2008 - 2010 are not directly comparable to previous years, but the prevalence was probably approximately the same in 2008 as in 2007 and gradually decreasing in 2009 and 2010.

Relevance of the findings in animals to findings in foodstuffs and to human cases

The overall occurrence of positive broiler flocks is low, but there is a large seasonal variation with a peak during the summer and autumn, and the surveillance programme is therefore covering that period of the year. Even though approximately 75% of the positive flocks are discovered before slaughter, and thereby subject to compulsory freezing or heat treatment, the number of *Campylobacter* positive broiler carcasses on the market during the summer can be considerable.

Table *Campylobacter* in animals

	Source of information	Sampling unit	Units tested	Total units positive for <i>Campylobacter</i> spp.	<i>C. jejuni</i>	<i>C. coli</i>	<i>C. upsaliensis</i>	<i>C. spp.</i> , unspecified
<i>Gallus gallus</i> (fowl)								
Broilers – at farm	NACB ¹	flock	2170	110				110
Cattle	NVI ²	animal	121	26	17	1		8
Sheep	NVI ²	animal	16	6	5			1
Dogs	NVI ²	animal	386	122	17	2	94	9
Cats	NVI ²	animal	97	9	3		4	2

¹ NACB = Norwegian Action plan against *Campylobacter* in Broilers. Only covering the peak season 1 May - 31 October. There is no data available on the *Campylobacter* species because the method used is a Real time PCR method where no isolates are obtained.

² NVI = Norwegian Veterinary Institute: Diagnostic samples.

Antimicrobial resistance in *Campylobacter* isolates

Antimicrobial resistance in *Campylobacter* sp. from cattle

Sampling strategy used in monitoring

Samples from healthy animals included in various surveys were investigated for occurrence of *Campylobacter* spp. One isolate per positive farm was included for susceptibility testing.

Type of specimen taken

Faecal samples collected at farm.

Procedures for the selection of isolates for antimicrobial testing

One isolate of *Campylobacter jejuni* from each positive holding was selected for antimicrobial testing.

Methods used for collecting data

Strains were isolated and tested for the antimicrobial susceptibility at the Norwegian Veterinary Institute in Oslo.

Laboratory methodology used for identification of the microbial isolates

NMKL No 119 without enrichment.

Laboratory methods used for detection for resistance

The VetMIC microdilution method (Dept. of Antibiotics, National Veterinary Institute, Sweden) was used for the susceptibility testing of all isolates. The antimicrobials included are listed in the table.

Breakpoints used in testing

Epidemiological cut-off values recommended by EFSA were used.

Results

The qualitative data are presented in the table. Quantitative data as well as data on breakpoints and range of testing are presented in the NORM/NORM-VET 2010 report.

Table Antimicrobial susceptibility testing of *Campylobacter jejuni*

	Cattle
Number of isolates tested	11
Number of isolates resistant to	
Tetracycline	0
Ciprofloxacin	1
Streptomycin	1
Gentamicin	0
Erythromycin	0
Nalidixic acid	1

Listeriosis

General evaluation of the national situation

History of the disease and/or infection in the country

Listeriosis is endemic in Norway with sporadic clinical cases in humans and animals, especially among sheep. Since 1982, the number of notified human cases has varied from 2-50. The incidence rate has varied from 0.05-1.07 per 100 000. Most of the cases are sporadic, occurring in old people or persons with an underlying disease. A few congenital cases have been reported.

An outbreak occurred in 1992 which involved six reported cases and was traced back to contaminated, vacuum packed cold cuts from a Norwegian meat producer. In 2005 a hospital outbreak occurred with 3 cases, probably linked to cold cuts (the same strain of *L. monocytogenes* as isolated from the patients was found on the slicing machine in the hospital kitchen). In 2007 another outbreak with 21 verified cases occurred and was caused by contaminated soft cheese.

In a survey conducted in 1994, the prevalence of *L. monocytogenes* in samples of vacuum packed cold cuts and smoked salmon was 1.7% and 7.8%, respectively. The prevalence in smoked salmon was 3.4% in a survey conducted in 1996-1997. In 2002 4.3% of 703 samples of domestically produced fish and fish products, mainly unprocessed and smoked salmon, were positive for *L. monocytogenes*. In 2003, 8.6% of 990 samples of smoked salmon taken at retail level were positive for *L. monocytogenes*. The level of contamination was less than 10 CFU/g in 53 samples, between 10 and 100 in 20 samples, between 100 and 1000 in 10 samples and more than 1000 CFU/g in two samples. In a survey conducted in 1995 involving ready-to-eat poultry products, the prevalence of *L. monocytogenes* was 0.4%. A survey of domestically produced raw milk products conducted in 1999 revealed that one out of 282 samples (0.4%) was positive for *L. monocytogenes*. A survey of raw bulk milk at Norwegian dairy farms, also conducted in 1999, did not detect any *L. monocytogenes* in 336 samples from cattle bulk milk, whereas four of 100 samples from goat bulk milk were positive for *L. monocytogenes*. This illustrates that products made of raw milk might be risk products with regard to *L. monocytogenes*.

Fermented trout is a traditional food product in Norway that is consumed without heat treatment. Studies have shown that fermented trout frequently is contaminated with *L. monocytogenes*, sometimes in high concentrations (up to 2000 CFU per gram). Former guidelines issued by the Food Safety Authority recommended a maximum level of 1000 CFU per gram for this particular product combined with information about risk products to vulnerable consumers. Recent studies have shown that it is possible to produce fermented trout without *L. monocytogenes* if hygienic precautionary measures, including temperature control and appropriate salt levels, are implemented throughout the process.

National evaluation of the recent situation, the trends and sources of infection

Listeriosis is endemic in Norway with sporadic clinical cases in animals, especially among sheep. However, listeriosis is not a common disease in humans in Norway. Most cases are sporadic and seen in the elderly or in patients with underlying disease. Ready-to-eat products have been identified as a source for human listeriosis.

Recent actions taken to control the zoonoses

Until the Regulation (EC) No 2073/2005 on microbiological criteria was implemented in March 2010, the Norwegian Food Safety Authority recommended that findings of *L. monocytogenes* in ready-to-eat food products with a shelf life longer than 15 days and in which the bacteria easily could grow, should result in recall from the market of the corresponding lot. Since then the requirements of the Regulation (EC) No 2073/2005 apply, i.e., monitoring of the production process, shelf-life studies when deemed appropriate, withdrawal from the market by unsatisfactory results and taking measures to prevent the recurrence of the contamination, such as reviewing the production routines and shelf life of the product. Dietary advice is given to pregnant women.

Listeriosis in humans

Reporting system in place for the human cases

Human cases are reported to the Norwegian Surveillance System for Communicable Diseases (MSIS), from microbiological laboratories as well as from clinical doctors. The system distinguishes between domestic and imported cases. The severity of the disease at the time of reporting is also recorded. However, the surveillance system does not follow individual patients over time to record further disease development and final outcome.

Case definition

A case from which *L. monocytogenes* has been detected in blood, cerebrospinal fluid or other normally sterile sites or a case with serology indicating recent infection.

Diagnostic/analytical methods used

Bacteriology (isolation of *L. monocytogenes* from a normally sterile site) followed by voluntary confirmation (species identification and serotyping) at the National Reference Laboratory.

Notification system in place

According to the Communicable Disease Act, human cases are notifiable to the Norwegian Surveillance System for Communicable Diseases (MSIS) since 1975.

History of the disease and/or infection in the country

Since 1982, the number of notified cases has varied from 2-50. The incidence rate has varied from 0.05-1.07 per 100 000. Most of the cases are sporadic, occurring in elderly individuals or persons with underlying disease. A few congenital cases are also being reported. The first recorded outbreak of listeriosis in Norway occurred in 1992, involving six reported cases. The outbreak was linked to vacuum packed cold cuts. In 2005, an outbreak occurred in a hospital in the middle of Norway. Three cases were reported, and the outbreak was linked to cold cuts. Another outbreak occurred in 2007, involving 21 reported cases of whom two died. The outbreak was linked to a Norwegian pasteurised soft cheese.

Results of the investigation

In 2010, a total of 23 confirmed cases of listeriosis were notified (incidence rate 0.5 per 100 000), 19 cases were infected in Norway, and one of the cases reported having been infected abroad (Sweden). One of the cases was pregnancy-associated. Two deaths were recorded.

National evaluation of the recent situation, the trends and sources of infection

Listeriosis in humans is a relatively rare disease in Norway and has been so for many years. Most of the cases are sporadic, occurring in elderly individuals or persons with underlying diseases. There is, however, an increasing trend if we look at the number of recorded cases over a twenty year period. The increase is not seen in pregnancy-associated cases. The reason for this increasing trend is unknown, but could be related both to an increase in the number of in elderly individuals and persons with other underlying diseases, and to increased exposure to *L. monocytogenes* in consumed food.

Relevance as zoonotic disease

Listeriosis in humans is a relatively rare disease in Norway.

Listeria in foodstuffs

Monitoring system

No continuous monitoring of foodstuffs takes place. Surveys are occasionally performed. Norway follows the EU requirements regarding testing for *L. monocytogenes* in ready-to-eat foods (Regulation (EC) NO 2073/2005). Samples are taken as part of internal control programmes in the food producing industry.

Definition of positive finding

A positive sample is a sample from which *Listeria* spp. has been isolated.

Diagnostic/analytical methods used

At the production plant: NMKL No 136:2007

At retail: NMKL No 136:2007, ISO 11290-1:1996/Amd 2004, ISO 11290-2:1998

Control program/mechanisms

No official control programmes in place. When relevant, monitoring and control take place as an integral part of food business operators' internal control systems.

Measures in case of the positive findings

Previously, the Norwegian Food Safety Authority recommends that findings of *L. monocytogenes* in ready-to-eat food products with a shelf life longer than 15 days and in which the bacteria could easily grow, should result in recall from the market of the corresponding lot. Since 1st March 2011 the requirements of the Regulation (EC) No 2073/2005 apply, i.e., monitoring of the production process, shelf-life studies when deemed appropriate, withdrawal from the market by unsatisfactory results and taking measures to prevent the recurrence of the contamination, such as reviewing the production routines and shelf life of the product.

Results of the investigation

In 2010, a total of 85 samples of fish from wild stock and 28 samples of farmed fish were investigated. A total of three samples of farmed fish were positive for *L. monocytogenes*, but in very low concentrations.

National evaluation of the recent situation, the trends and sources of infection

In general, the occurrence of *L. monocytogenes* in food products is low.

Listeria in animals

Monitoring system

Listeriosis is a notifiable disease in animals. There are no active surveillance regarding *L. monocytogenes* in animals. Information is achieved through clinical and laboratory reports.

Frequency of the sampling

When there is a suspected case.

Case definition

A case may be defined by 1) positive histopathology combined with clinical signs, 2) positive bacteriology.

Diagnostic/analytical methods used

Bacteriology, histopathology and immunohistochemistry.

Measures in case of the positive findings or single cases

Normally none.

Notification system in place

Listeriosis has been a list C disease according to the Animal Disease Act since 1965.

Results of the investigation

Many animals are investigated with regard to *L. monocytogenes* and listeriosis in clinical laboratories. In 2010, at the Norwegian Veterinary Institute, 35 sheep, 25 goats, four cattle and one alpaca were found positive.

Relevance of the findings in animals to findings in foodstuffs and to human cases

Listeria spp. is present in the environment and also in food producing animals. However, there is no epidemiological evidence that listeriosis in humans are linked to listeriosis in animals.

E. coli infections

General evaluation of the national situation

History of the disease and/or infection in the country

The reported incidence of VTEC infections in humans in Norway has so far been low. Approximately half of the cases are acquired domestically. In 2006 there was a severe outbreak caused by VTEC O103:H25 with 17 patients, out of which 10 developed HUS and one died. In 2009 there was another outbreak, caused by sorbitol fermenting *E. coli* O157:H-. There were 19 cases, out of which 9 developed HUS and one died. In 2010, another outbreak occurred, with 3 cases infected with the same strain as in the 2009-outbreak.

A study conducted in 1995 revealed a low prevalence of VTEC O157 among Norwegian dairy cattle; animal prevalence 0.3% and herd prevalence 1.0%. In a survey conducted in 1998-1999, one out of 574 dairy cattle herds were positive for VTEC O157 (herd prevalence 0.2%, animal prevalence between 0.02 and 0.06%). In 2000, none of the tested 1435 beef cattle from 165 herds were positive for VTEC O157. A survey in 2002, in which 453 pooled faecal samples from 155 beef cattle herds were tested for the presence of VTEC O26, O103, O111, O145 and O157, revealed five pooled samples from five herds positive for VTEC O103, all *eae* negative.

In the surveillance programme for VTEC O157 in cattle, sheep, and goat carcasses running in the period 1998-2004, the total carcass prevalence was 0.06% for cattle and 0.03% for sheep. None of the 510 goat carcasses tested were positive.

In a national survey in sheep conducted in 2006-2007, samples from 585 flocks were analysed, 94 flocks from 2006 and 491 flocks from 2007. VTEC O103:H2 (*stx*₁ and *eae* positives) were detected in 0.7% and VTEC O157:H7 (*stx*₂ and *eae* positives - one was also *stx*₁ positive) in 0.9% of the flocks. Only the 2007 samples were analysed for *E. coli* O26, and VTEC O26 were detected in 0.8% of the flocks. In addition *stx* negative and *eae* positive *E. coli* O26 were detected in 16.1%, *stx* negative and *eae* positive *E. coli* O103:H2 in 3.1%, and *stx* negative and *eae* positive *E. coli* O103:H25 in 5.8% of the flocks.

National evaluation of the recent situation, the trends and sources of infection

Although the annual incidence in humans in Norway up to 2006 was low and predominantly involved sporadic cases, the fear that the incidence might increase in the future, and that outbreaks may occur proved valid in 2006. Data show that VTEC O157 is present in the cattle and sheep populations, and although the prevalences seem to be low, this reservoir represents a source of possible human infection. The 2006 outbreak caused by VTEC O103:H25 showed that other VTEC than the "high five" (VTEC O26, O103:H2, O111, O145 and O157) may be of potential danger to humans.

Relevance of the findings in animals, feedingstuffs and foodstuffs to human cases

Although the prevalence of VTEC O157 in the cattle and sheep populations seems to be low, there are other VTEC where the knowledge is sparse. In general, there is always a potential for contamination in the food chain, which requires alertness at all steps from primary production, through processing, and retail and food preparation, as well as alertness among physicians and diagnostic laboratories.

E. coli infections in humans

Reporting system in place for the human cases

Human cases are reported to the Norwegian Surveillance System for Communicable Diseases (MSIS), from microbiological laboratories as well as from clinical doctors. The system distinguishes between domestic and imported cases. The severity of the disease at the time of reporting is also recorded. However, the surveillance system does not follow individual patients over time to record further disease development and final outcome. Haemolytic uremic syndrome (HUS) became a notifiable disease in December 2006. Before that, HUS was not notifiable per se, but was reported in relation to an EHEC diagnosis.

Case definition

A case from which enterohaemorrhagic *E. coli* or its toxins have been detected from faecal samples.

Diagnostic/analytical methods used

Clinical microbiological laboratories use plating on selective media (such as SMAC) in order to detect presumptive VTEC O157 and/or genetic methods directed towards detection of Shiga toxin genes followed by isolation of VTEC and confirmation at the National Reference Laboratory. Confirmation includes examination for the presence of Shiga toxin genes and other virulence factors.

Notification system in place

According to the Communicable Disease Act, human cases are notifiable to the Norwegian Surveillance System for Communicable Diseases (MSIS) since 1995. Haemolytic uremic syndrome (HUS) became a notifiable disease in December 2006.

History of the disease and/or infection in the country

The reported incidence of VTEC infections in humans in Norway has been low. The number of cases has varied between 0-51 per year, except in 2009 with 108 reported cases. Approximately half of the cases are acquired domestically. Most reported cases are caused by VTEC O157.

The first foodborne VTEC outbreak in Norway occurred in 1999 and involved four culture positive patients (O157). Epidemiological investigations incriminated domestically produced lettuce as the most likely source of infection. A severe outbreak caused by VTEC O103:H25 in 2006 involved 17 patients of which 10 developed HUS and one died. In 2009, an outbreak caused by sorbitol fermenting O157:H- occurred. Thirteen children got ill, and of these nine developed HUS and one child died. The source of the outbreak was not found.

Results of the investigation

In 2010, 51 cases (incidence rate 1.0 per 100.000) of VTEC and HUS were reported. A total of 5 cases of HUS were reported; sorbitol fermenting O157:H- was isolated from 3 of the patients, O121:H? from one patient, and O26:H11 from one patient. Of the 51 cases of VTEC infections reported, the most commonly isolated serotypes were O103 (11 cases), O157 (8 cases) and O26 (2 cases). Eleven of the 51 cases reported contracting the infection abroad. There was one outbreak with HUS counting 3 cases.

National evaluation of the recent situation, the trends and sources of infection

The number of cases reported in 2010 is much lower than the cases reported in 2009. One outbreak counting three HUS cases was reported, as opposed to the seven outbreaks reported last year. Many of the notified cases in 2009 were detected because of increased attention and testing due to the outbreaks. The laboratory methods have probably improved since the O103 outbreak Norway experienced in 2006. Even though there is a decrease from 2009 to 2010, the 51 cases of VTEC and 5 cases of HUS reported in 2010 reflects an increasing trend for EHEC in Norway. The reason for this increase is unknown.

Relevance as zoonotic disease

Data show that VTEC is present in the cattle and sheep populations, although the prevalences seem to be low. However, there is also a reservoir of *E. coli* with *eae*, but *stx* negative, that may be of concern as human pathogenics (aEPEC) or as precursors for VTEC. Thus, there is a potential for contamination in the food chain or by direct animal contact, which requires alertness at all steps from primary production, through processing, and retail and food preparation, as well as alertness among physicians and diagnostic laboratories.

Additional information

Patients whose work represents a risk for spread of the disease, e.g., people working with food production, children in day-care and health care personell, are advised to stay away from work while they have symptoms. It is recommended that for these patients five consecutive faecal samples examined after the symptoms have disappeared should be negative before returning to work.

Verotoxigenic E. coli (VTEC) in animals

Monitoring system

Prevalence surveys in cattle, sheep and goats have been conducted occasionally since 1998. In 2006-2007 a survey regarding VTEC in sheep was conducted, with a total of 593 flocks sampled. Single faecal samples were collected from the 50 youngest animals in each flock.

Type of specimen taken

Animals at farm: Faeces

Case definition

An animal or herd from which VTEC is isolated.

Diagnostic/analytical methods used

Modification of NMKL No 164:1999 with IMS (or IMS-ELISA) followed by virulence characterization by PCR.

Measures in case of the positive findings or single cases

If VTEC O157 or other VTEC that can pose a health risk for humans is detected in an official survey among live animals, the Norwegian Food Safety Authority and Municipal Medical Officer are notified. Restrictions may be imposed on livestock holdings where such VTEC is detected. The holdings sampled in the survey of sheep flocks in 2006-2007 were anonymized.

Notification system in place

Findings in carcasses of VTEC O157 or other VTEC that can pose a health risk to humans lead to condemnation of the carcasses and notification to the authorities. Findings of such VTEC in samples from live animals are not notifiable as an animal disease, but since VTEC is a pathogen that can be transmitted from animals to humans, competent authorities have to be informed about positive findings.

Results of the investigation

In 2010 more than 200 samples, mainly from food but also some from animals and environment were investigated at the Norwegian Veterinary Institute due to follow up of human cases. Definite links between human cases and the samples from food, animals and environment were not identified except for one case in a young child where the same O26 strain was found in faeces from calves at a petting zoo the child had visited.

National evaluation of the recent situation, the trends and sources of infection

The prevalence of human pathogenic VTEC O157, O103, O26, O45 and O111 is still considered low in Norwegian cattle, sheep and goats.

Tuberculosis, mycobacterial diseases

General evaluation of the national situation

History of the disease and/or infection in the country

Norway has been granted the officially tuberculosis-free status of bovine herds by the EFTA Surveillance Authority (ESA) (EFTA Surveillance Authority Decision No 28/07/COL) as Norway fulfils the requirements laid down in Council Directive 64/432/EEC as amended. Bovine tuberculosis (*M. bovis*) was declared eliminated in cattle in Norway in 1963 as a result of an official eradication programme against the disease. During the period 1895-1896, 26% of 2195 tuberculin-tested herds were positive. In 1950, 18 herds were registered as being infected, while in the beginning of the 1960s only one or two infected herds were reported annually. Since bovine tuberculosis was declared eliminated, it has only been recorded three times; in 1984 in two cattle herds and in 1986 in one cattle herd. These herds were in the same geographical area and the origin of the infection in these herds was probably a man with tuberculosis. Tuberculosis caused by *M. bovis* in other animal species than cattle has not been recorded in Norway after the disease was eliminated from cattle in 1963. Tuberculosis in humans caused by *M. bovis* is only sporadically recorded in Norway, and since 1977 the few recorded cases have been imported except for one case of reactivation in 1994.

National evaluation of the recent situation, the trends and sources of infection

As Norway is officially free from bovine tuberculosis, the probability of contracting *M. bovis* infection from Norwegian animals or animal products of Norwegian origin is close to zero.

Relevance of the findings in animals, feedingstuffs and foodstuffs to human cases

There have been no findings of *M. bovis* in animals or foodstuffs. The probability of contracting *M. bovis* infection from Norwegian animals or animal products of Norwegian origin is close to zero.

Tuberculosis due to Mycobacterium bovis in humans

Reporting system in place for the human cases

Human cases are reported to the Norwegian Surveillance System for Communicable Diseases (MSIS), from microbiological laboratories as well as from clinical doctors. The system distinguishes between Norwegian and foreign born cases. The severity of the disease at the time of reporting is also recorded. The surveillance system includes individual treatment outcome data for all tuberculosis patients.

Case definition

A confirmed case of *M. bovis*, *M. tuberculosis*, or *M. africanum* is a case that has been confirmed by isolation of *M. bovis*, *M. tuberculosis*, or *M. africanum*, respectively. Cases of tuberculosis that are diagnosed without laboratory confirmation (diagnoses based on clinical symptoms and X-ray examination) are also notified and included in the statistics.

Diagnostic/analytical methods used

Clinical indications: Bacteriology, X-ray, pathology.

Screening: Miniature X-ray, tuberculin skin testing, Interferon gamma release assays.

Notification system in place

According to the Communicable Disease Act, human cases caused by bacilli belonging to the *M. tuberculosis* complex (including *M. tuberculosis*, *M. bovis*, and *M. africanum*) are notifiable to the Norwegian Surveillance System for Communicable Diseases (MSIS) since 1975, and before that notifiable to a separate Tuberculosis Register since 1900.

History of the disease and/or infection in the country

The incidence of human tuberculosis (*M. bovis* and *M. tuberculosis*) has steadily decreased during the last 50 years in persons of Norwegian origin. BCG vaccination was introduced in 1947 and was mandatory until 1995. Pasteurisation of milk for commercial sale became mandatory in 1951. Since 1977, the annual incidence rate in persons born in Norway has decreased from 11 to 1.4 per 100 000, and most cases in this part of the population are recurrent cases in elderly patients. Along with increased immigration to Norway, the proportion of tuberculosis cases involving persons born outside Norway has increased during the last two decades (from less than 10% in 1977 to 81% in 2006). Since bovine tuberculosis in cattle was eliminated in Norway in 1963, almost all bacteriologically confirmed cases in humans have been caused by *M. tuberculosis*. The last domestic case of tuberculosis caused by *M. bovis* was reported in 1994 in a 100 year old woman infected in her youth. Apart from this case, no indigenous cases of tuberculosis caused by *M. bovis* in humans have been reported since 1977. Imported cases of tuberculosis caused by *M. bovis* are sporadically reported; in 2010 in one patient from Africa, in 2005 in two patients from Somalia and Afghanistan, respectively, in 2002 one patient from Somalia, in 2001 one patient from Tanzania, in 2000 two patients from Somalia and Morocco, respectively, in 1999 one patient from Sri Lanka, in 1998 one patient from Somalia, and in 1994 one patient infected in India.

Results of the investigation

In 2010, one case with tuberculosis caused by *M. bovis* was notified. The patient was infected in Africa.

National evaluation of the recent situation, the trends and sources of infection

Tuberculosis caused by *M. bovis* is only sporadically recorded in Norway, and except for a case of reactivation in 1994, the few recorded cases reported since 1977 have been imported.

Relevance as zoonotic disease

As Norway is officially free from bovine tuberculosis, the probability of contracting *M. bovis* infection from Norwegian animals or animal products of Norwegian origin is close to zero.

Additional information

In Norway, the child vaccination programme has included vaccination against tuberculosis since 1947. The BCG vaccine (live attenuated *M. bovis*) is offered to unvaccinated and tuberculin negative persons belonging to certain risk groups; immigrants from countries with high prevalence of tuberculosis, persons travelling to high endemic areas for a prolonged time period, teachers, health personnel, personnel on ships and in offshore industry, and military personnel. In addition, the BCG vaccine is offered to all children during junior high school (13-14 years old). In general, the immunisation coverage in Norwegian children is high; for the BCG vaccine it is estimated to be 99%. In Norway, the BCG vaccine is estimated to give 80% protection against tuberculosis. Tuberculin skin test is mandatory for immigrants coming to Norway from high prevalence countries. Immigrants who are 15 years or older must also undergo chest radiograph screening. Screening for tuberculosis in certain risk populations is sometimes conducted.

Mycobacterium in animals

A. *Mycobacterium bovis* in bovine animals

Status as officially free of bovine tuberculosis during the reporting year: The entire country free Norway has been granted the officially tuberculosis free status of bovine herds by the EFTA Surveillance Authority (ESA) (EFTA Surveillance Authority Decision No 28/07/COL) as Norway fulfils the requirements laid down in Council Directive 64/432/EEC as amended.

Monitoring system

Every slaughtered animal, except animals slaughtered for on-the-farm consumption, is subjected to meat inspection regarding tuberculosis (lymph node examination) according to Regulation (EC) No 854/2004 (until the 1st of March: Council Directive 64/433/EEC). All breeding bulls are tuberculin tested several times. Imported animals are tuberculin tested if considered relevant based upon individual assessment. If suspicion arises whether an animal may have tuberculosis (sick or dead animal), relevant tests will be carried out.

Frequency of the sampling

All slaughtered animals are subject to meat inspection. Imported animals are tested during week 22 of the six months long isolation period. Breeding bulls are tuberculin tested before being transferred to a semen collection centre and thereafter subject to yearly testing.

Type of specimen taken

Animals for slaughter: Lymph nodes. Breeding animals and imported animals: Tuberculin testing.

Methods of sampling (description of sampling techniques)

Slaughtered animals: Meat inspection at the slaughterhouse; lymph node examination. Imported animals and breeding animals: Tuberculin testing. Clinical indication: Methods will vary depending on the problem.

Case definition

A single animal from which *M. bovis* or *M. tuberculosis* has been isolated. Herd is the epidemiological unit.

Diagnostic/analytical methods used

Slaughtered animals: Meat inspection regarding tuberculosis (lymph node examination) according to Regulation (EC) No 854/2004 (until the 1st of March: Council Directive 64/433/EEC). If indicated: bacteriology and histology. Clinical indications: Tuberculin testing (intradermal comparative test), pathology, and/or bacteriology. Breeding animals and imported animals: Tuberculin testing (intradermal comparative test).

Vaccination policy

Vaccination of animals against tuberculosis is prohibited in Norway.

Control program/mechanisms

Every slaughtered animal, except animals slaughtered for on-the-farm consumption, is subjected to meat inspection regarding tuberculosis (lymph node examination) according to Regulation (EC) No 854/2004 (until the 1st of March: Council Directive 64/433/EEC).

Measures in case of the positive findings or single cases

Norway would as a minimum implement the measures as laid down in Council Directive 64/432/EEC as amended in case of positive findings or if suspicion of tuberculosis in bovine animals should arise.

Notification system in place

Tuberculosis caused by *M. bovis* or *M. tuberculosis* of all species has been a notifiable List B disease according to the Animal Diseases Act since 1894.

Results of the investigation

In 2010, two slaughtered bovine animals had findings at slaughter indicating tuberculosis, and was submitted for examination for *Mycobacterium* sp. The samples were negative.

A total of 275 bulls owned by a breeding company all had negative tuberculin tests.

National evaluation of the recent situation, the trends and sources of infection

Bovine tuberculosis was declared eliminated in cattle in 1963.

Relevance of the findings in animals to findings in foodstuffs and to human cases

There have been no findings of *M. bovis* in animals or foodstuffs. The risk for humans contracting tuberculosis from livestock within the country is negligible.

B. *Mycobacterium bovis* in farmed deer

Monitoring system

Every slaughtered animal, except animals slaughtered for on-the-farm consumption, is subjected to meat inspection regarding tuberculosis (lymph node examination) by an official veterinarian according to Regulation (EC) No 854/2004 (until the 1st of March: Council Directive 64/433/EEC) Imported deer are tuberculin tested if considered relevant based upon individual assessment. If suspicion arises whether an animal may have tuberculosis (sick or dead animal), relevant tests will be carried out.

Frequency of the sampling

All slaughtered animals are subject to meat inspection. Imported deer are tested during week 5 of the two months long isolation period.

Type of specimen taken

Animals for slaughter: Lymph nodes. Imported animals: Tuberculin testing.

Methods of sampling (description of sampling techniques)

Slaughtered animals: Meat inspection at the slaughterhouse; lymph node examination. Imported animals: Tuberculin testing. Clinical indications: Methods will vary depending on the problem.

Case definition

A single animal from which *M. bovis* or *M. tuberculosis* has been isolated. The herd is the epidemiological unit.

Diagnostic/analytical methods used

Slaughtered animals: Meat inspection regarding tuberculosis (lymph node examination) by an official veterinarian according to Regulation (EC) No 854/2004 (until the 1st of March: Council Directive 64/433/EEC). If indicated: bacteriology and histology. Imported animals: Tuberculin testing (intradermal comparative test). Clinical indications: Tuberculin testing (intradermal comparative test), pathology, and/or bacteriology.

Vaccination policy

Vaccination of animals against tuberculosis is prohibited in Norway.

Control program/mechanisms

Every slaughtered animal, except animals slaughtered for on-the-farm consumption, is subjected to meat inspection regarding tuberculosis (lymph node examination) by an official veterinarian according to Regulation (EC) No 854/2004 (until the 1st of March: Council Directive 64/433/EEC). Required autopsy of animals older than 12 months of age that die or are killed because of a disease.

Measures in case of the positive findings or single cases

Norway would as a minimum implement the measures as laid down in Council Directive 64/432/EEC as amended in case of positive findings or if suspicion of tuberculosis should arise.

Notification system in place

Tuberculosis caused by *M. bovis* or *M. tuberculosis* of all species has been a notifiable List B disease according to the Animal Diseases Act since 1894.

Results of the investigation

In 2010, material from one farmed deer was submitted for examination for *Mycobacterium* sp. The sample was negative.

National evaluation of the recent situation, the trends and sources of infection

Bovine tuberculosis has not been diagnosed in farmed deer in Norway. The population of farmed deer is very small in Norway.

Relevance of the findings in animals to findings in foodstuffs and to human cases

There have been no findings of *M. bovis* in animals or foodstuffs. The risk for humans contracting tuberculosis from livestock within the country is negligible.

C. *Mycobacterium* spp. in other animals

Monitoring system

For cattle and farmed deer, see the respective chapters. Every slaughtered animal, except poultry and animals slaughtered for on-the-farm consumption, is subjected to meat inspection regarding tuberculosis (lymph node examination) by an official veterinarian according to Regulation (EC) No 854/2004 (until the 1st of March: Council Directive 64/433/EEC). Imported animals are tuberculin tested if considered relevant based upon individual assessment. Boars selected for export of semen to USA are tuberculin tested. If suspicion arises whether an animal may have tuberculosis (sick or dead animal), relevant tests will be done.

Frequency of the sampling

All slaughtered animals are subject to meat inspection. Imported animals: Sheep and goats are tested during week 23 of the two years long isolation period. Pigs are tested during week 7 of the two months long isolation period. Lamas are tested during week 22 of the six months long isolation period.

Type of specimen taken

Animals for slaughter: Lymph nodes. Imported or exported animals: Tuberculin testing.

Methods of sampling (description of sampling techniques)

Slaughtered animals: Meat inspection at the slaughterhouse; lymph node examination. Imported animals: Tuberculin testing. Clinical indications: Methods will vary depending on the problem.

Case definition

A single animal from which *M. bovis* or *M. tuberculosis* has been isolated. The herd is the epidemiological unit.

Diagnostic/analytical methods used

Slaughtered animals: Meat inspection regarding tuberculosis (lymph node examination) by an official veterinarian according to Regulation (EC) No 854/2004 (until the 1st of March: Council Directive 64/433/EEC). If indicated: bacteriology and histology. Tests of imports, exports: Tuberculin testing (intradermal comparative test). Clinical indications: Tuberculin testing (intradermal comparative test), pathology, and/or bacteriology.

Vaccination policy

Vaccination of animals against tuberculosis is prohibited.

Control program/mechanisms

Every slaughtered animal, except poultry and animals slaughtered for on-the-farm consumption, is subjected to meat inspection regarding tuberculosis (lymph node examination) by an official veterinarian according to Regulation (EC) No 854/2004 (until the 1st of March: Council Directive 64/433/EEC).

Measures in case of the positive findings or single cases

Norway would as a minimum implement the measures as laid down in Council Directive 64/432/EEC as amended in case of positive findings or if suspicion of tuberculosis should arise.

Notification system in place

Tuberculosis caused by *M. bovis* or *M. tuberculosis* in all species has been a notifiable List B disease according to the Animal Diseases Act since 1894. Cases are to be notified to the Norwegian Food Safety Authority.

Results of the investigation

In 2010, tuberculin tests were performed on 99 breeding boars at AI stations, all were negative. Samples from four pigs, three dogs, one sheep and one horse were analyzed for the presence of *Mycobacterium* species. The animals were negative.

National evaluation of the recent situation, the trends and sources of infection

Bovine tuberculosis was declared eliminated in cattle in 1963, and has since then not been recorded in other animal species.

Relevance of the findings in animals to findings in foodstuffs and to human cases

There have been no findings of *M. bovis* in animals or foodstuffs. The risk for humans contracting tuberculosis from livestock within the country is negligible.

Brucellosis

General evaluation of the national situation

History of the disease and/or infection in the country

Bovine brucellosis has been a notifiable disease since 1903. An offensive eradication programme to eliminate the disease was launched in 1935, and Norway was declared free from bovine brucellosis in 1953. Ovine, caprine, or porcine brucellosis has never been recorded in Norway. Norway has been granted official brucellosis-free status of bovine herds by the EFTA Surveillance Authority (ESA) (EFTA Surveillance Authority Decision No 28/07/COL). Also regarding *Brucella melitensis*, Norway fulfils the requirements for an officially free status for the disease in sheep and goats, however, a formal decision is not available. Human brucellosis has always been a rare disease in Norway, the majority of the cases being imported, and a few cases due to laboratory infections domestically.

National evaluation of the recent situation, the trends and sources of infection

As bovine brucellosis was declared eliminated in Norway in 1953, and ovine, caprine, or porcine brucellosis has never been recorded, Norway is considered free from brucellosis in production animals. Research studies have shown that antibodies against *Brucella* can be detected in marine mammals (minke whales and hooded seals) from the North Atlantic Ocean, and in polar bears from the archipelago of Svalbard and the Barents Sea. *Brucella* sp. different from previously described species has also been isolated from hooded seals from the Greenland Sea.

Relevance of the findings in animals, feedingstuffs and foodstuffs to human cases

There have been no findings of *Brucella* spp. in terrestrial animals or foodstuffs. The probability of contracting brucellosis from Norwegian animals or animal products of Norwegian origin is close to zero.

Brucellosis in humans

Reporting system in place for the human cases

Human cases are reported to the Norwegian Surveillance System for Communicable Diseases (MSIS), from microbiological laboratories as well as from clinical doctors. The system distinguishes between domestic and imported cases. The severity of the disease at the time of reporting is also recorded. However, the surveillance system does not follow individual patients over time to record further disease development and final outcome.

Case definition

A clinically compatible case that is laboratory confirmed.

Diagnostic/analytical methods used

Serology (serum antibody test or antigen test of clinical specimen) and bacteriology (isolation).

Notification system in place

According to the Communicable Disease Act, human cases are notifiable to the Norwegian Surveillance System for Communicable Diseases (MSIS) since 1975.

History of the disease and/or infection in the country

Human brucellosis has always been a rare disease in Norway. During the period 1983-2010, only 20 cases of brucellosis were reported: In 2010 two cases infected abroad. In 2006 three cases of which two had travelled to countries outside Europe and for the third case, there was no information available. In 2005 one case infected in Africa. In 2004 two cases; one infected at work (health care/laboratory), the other infected in Cyprus. In 2003 three cases; two probably infected in Ethiopia and one probably infected in a laboratory. In 2002 three cases; from Spain, Iraq and Georgia. In 2001 two cases; both probably infected in Lebanon. In 2000 one case infected in Turkey probably through milk. In 1999 one case infected through milk in Turkey. In 1997 one immigrant from Turkey. In 1987 a Norwegian UN soldier stationed in Lebanon (*B. melitensis*).

Results of the investigation

In 2010 two cases were reported. Both cases were infected abroad.

National evaluation of the recent situation, the trends and sources of infection

Brucellosis is rarely recorded in Norway. Since 1983, only 18 cases have been recorded. Two of these are known to be infected in Norway, both laboratory contracted.

Relevance as zoonotic disease

As Norway is free from brucellosis in terrestrial food producing animals, the risk of humans contracting brucellosis from such animals or from Norwegian animal products is considered negligible. However, the recent findings of *Brucella* species in marine mammals needs further research to better understand the epidemiology and to address possible public health implications.

Brucella in animals

A. *Brucella abortus* in bovine animals

Status as officially free of bovine brucellosis during the reporting year: The entire country free

Norway is regarded as officially free from bovine brucellosis according to the EFTA Surveillance Authority (ESA) (EFTA Surveillance Authority Decision No 28/07/COL).

Monitoring system

Surveillance programme: During the years 2000-2004, the programme consisted of an active surveillance part, where 20% of the Norwegian cattle population were sampled each year, and a passive surveillance part, where aborted fetuses and blood samples from their dams were investigated. Since 20% of the Norwegian cattle population had been tested annually for five consecutive years and thereby fulfilled the requirements from the EU, the programme in 2005 was reduced to passive surveillance only. According to the programme, all abortions between the fifth month of pregnancy and 14 days before expected birth in a herd in which there has been at least two such abortions the last 12 months, should be sampled. In addition, blood samples from the cow should be examined. All breeding bulls are tested. Imported animals are serologically tested if considered relevant, based upon an assessment of the health status in the country of origin. Tests are also carried out in connection with clinical indications and export.

Frequency of the sampling

All breeding bulls are tested serologically twice before being transferred to a semen collection centre, and subsequently retested within 12 months. Bulls are thereafter subject to yearly testing. Imported cattle are tested at week 22 during the six months long isolation period.

Type of specimen taken

Blood or foetus.

Methods of sampling (description of sampling techniques)

Surveillance programme: Foetus and the foetal membranes and paired blood samples from the mother are collected. Other monitoring systems: Blood samples. All samples are collected at farm.

Case definition

An animal which is seropositive for *Brucella* spp. even after retesting at least four weeks later, or an animal from which *Brucella* spp. has been isolated. The herd is the epidemiological unit.

Diagnostic/analytical methods used

Foetus: Full autopsy, histopathology, bacteriology. Blood samples from cows: Antibodies against *Brucella* in an indirect ELISA (Svanova). If the results are doubtful or positive, the samples are retested in duplicates. If the result still is doubtful or positive, the sample is tested with a competitive ELISA (C-ELISA, Svanova). If still positive, a complement fixation (CF) test is used. If the CF test is positive, new samples are taken four to six weeks after the initial sampling. If this is positive, or if there is a need for immediate follow up, the animal will be tested with an intracutane test using Brucellergene OCB from *B. melitensis* (Synbiotics). Breeding animals, imports, exports: Serology (Rose Bengal plate agglutination test, serum agglutination test or complement fixation test depending on the customer's demands). All tests are performed according to the OIE Manual of Diagnostic Tests and Vaccines for Terrestrial Animals, 5th ed. 2004. The indirect ELISA is standardized against EU Directive 64/432/EEC Annex C.

Vaccination policy

Vaccination of animals against brucellosis is prohibited in Norway.

Control program/mechanisms

The surveillance programme in cattle herds (in accordance to Council Directive 64/432/EEC Annex I) was established in 2000. All breeding bulls are serologically tested twice before being transferred to a semen collection centre, and subsequently within 12 months. Bulls are thereafter subjected to yearly testing. Imported cattle are serologically tested if considered relevant based upon an individual assessment. Tests are also carried out in connection with clinical indications and export.

Measures in case of the positive findings or single cases

Norway would as a minimum implement the measures as laid down in Council Directive 64/432/EEC as amended in case of positive findings or if suspicion of brucellosis in bovine animals should arise.

Notification system in place

Bovine brucellosis has been a notifiable List A disease according to the Animal Diseases Act since 1903. Cases are to be notified to the Norwegian Food Safety Authority.

Results of the investigation

In 2010, animals from a total of four herds were investigated in the surveillance programme (blood samples, aborted fetuses and bulk milk). A total of 491 bulls owned by a breeding company were tested for brucellosis. All samples were negative.

National evaluation of the recent situation, the trends and sources of infection

Bovine brucellosis was eliminated from Norway in 1953. No positive cases have been found since then.

Relevance of the findings in animals to findings in foodstuffs and to human cases

There have been no findings of *Brucella* spp. in cattle or foodstuffs from cattle. The probability of contracting brucellosis from Norwegian animals or animal products of Norwegian origin is close to zero.

Table *Brucella* spp. in animals

	Source of information ¹	Sampling unit	Units tested	Total units positive for <i>Brucella</i> spp.
Cattle	Surveillance programme	Herd	4	0
	BC	Animal	491	0
Sheep	Surveillance programme	Animal	8160	0
	BC	Animal	88	0
	NVI	Animal	18	0
Goats	Surveillance programme	Animal	779	0
Pigs	BC	Animal	1168	0
Dogs	NVI	Animal	27	0
Alpacas	NVI	Animal	14	0

¹ BC = Breeding Company, NVI=Norwegian Veterinary Institute (mainly tested in relation to export or import)

B. *Brucella melitensis* in sheep and goats

Status as officially free of ovine brucellosis during the reporting year: The entire country free

Due to its history in regard to *Brucella melitensis*, Norway fulfils the requirements for an officially free status for the disease.

Monitoring system

Surveillance programme sheep: A large proportion of herds being part of the breeding system with ram circles are tested. Randomly selected flocks not being part of any ram circles are also tested.

Surveillance programme goats: A large proportion of herds are selected for sampling each year. The programme started in 2007.

Imported sheep and goats are serologically tested if considered relevant based upon an assessment of the health status in the country of origin.

Frequency of the sampling

Surveillance programme: A selection of herds in the population is tested every year. Imported sheep and goats are tested for brucellosis at week 2 and 23 during the two year isolation period.

Type of specimen taken

Blood

Methods of sampling (description of sampling techniques)

Individual blood samples are collected at farm. Surveillance programme: In flocks with less than 30 animals, all animals are sampled; in herds with 30-100 animals, 30 are sampled; in herds with 100-200 animals, 35 are sampled; in herds with more than 200 animals, 40 animals are sampled.

Case definition

An animal which is seropositive for *Brucella* spp. or an animal from which *Brucella* spp. has been isolated. The herd is the epidemiological unit.

Diagnostic/analytical methods used

Rose Bengal plate agglutination test is used for the initial screening. A competitive ELISA (C-ELISA, Svanova) is used to follow up unclear or positive reactions due to possible cross reactions.

Vaccination policy

Vaccination of animals against brucellosis is prohibited.

Control program/mechanisms

The national surveillance programme and the control of imported animals are run by the Norwegian Food Safety Authority.

Measures in case of the positive findings or single cases

Norway would as a minimum implement the measures as laid down in Council Directive 91/68/EEC in case of positive findings or if suspicion of brucellosis in ovine or caprine animals should arise.

Notification system in place

Brucellosis in all species has been a notifiable List A disease according to the Animal Diseases Act since 1903. Cases are to be notified to the Norwegian Food Safety Authority.

Results of the investigation

Sheep: In 2010, in the surveillance programme, 8160 animals from 269 herds were tested for antibodies against *B. melitensis*. All were negative. All 88 rams tested for brucellosis were negative. All 18 animals tested in relation to export were negative.

Goats: In 2010, in the surveillance programme, 779 animals from 25 herds were tested for antibodies against *B. melitensis*. All were negative.

National evaluation of the recent situation, the trends and sources of infection

Ovine or caprine brucellosis has never been recorded in Norway.

Relevance of the findings in animals to findings in foodstuffs and to human cases

There have been no findings of *Brucella* spp. in sheep or goats or foodstuffs from sheep or goats. The probability of contracting brucellosis from Norwegian animals or animal products of Norwegian origin is close to zero.

D. *Brucella* spp. in pigs

Monitoring system

All breeding boars are tested. Imported pigs are tested if considered relevant based upon an individual assessment.

Frequency of the sampling

All breeding boars are tested twice before being transferred to a semen collection centre, and subsequently within 12 months or before slaughter. Imported pigs are tested during week 4 of the two months long isolation period.

Type of specimen taken

Blood

Methods of sampling (description of sampling techniques)

Blood samples are taken at the farms.

Case definition An animal which is seropositive for *Brucella* spp. or an animal from which *Brucella* spp. has been isolated. The herd is the epidemiological unit.

Diagnostic/analytical methods used

Rose Bengal plate agglutination test performed according to the latest edition of the OIE Manual of Diagnostic Tests and Vaccines for Terrestrial Animals, 2.8.5 Swine.

Vaccination policy

Vaccination of animals against brucellosis is prohibited in Norway.

Control program/mechanisms

All breeding boars are tested. Imported pigs are tested if considered relevant based upon an individual assessment.

Measures in case of the positive findings or single cases

If *Brucella* should be detected, the competent authorities must be notified without delay. Actions would be taken to identify and eliminate the source of the contamination in order to prevent further spread. Stringent restrictions including cleaning and disinfection, control of animal movement and control of person admission would be imposed on the infected holding. The whole herd would be destroyed.

Notification system in place

Brucellosis in all species has been a notifiable List A disease according to the Animal Diseases Act since 1903. Cases are to be notified to the Norwegian Food Safety Authority.

Results of the investigation

In 2010, all 1168 investigated pigs belonging to a breeding company tested negative. A total of 168 of these were tested in relation to export of live animals.

National evaluation of the recent situation, the trends and sources of infection

Porcine brucellosis has never been recorded in Norway.

Relevance of the findings in animals to findings in foodstuffs and to human cases

There have been no findings of *Brucella* spp. in swine or foodstuffs from swine. The probability of contracting brucellosis from Norwegian animals or animal products of Norwegian origin is close to zero.

Yersiniosis

General evaluation of the national situation

History of the disease and/or infection in the country

In the years 1982-1994, the number of notified cases in humans varied between 154 and 274 (mean 187). From 1994 there was a steady decline in the reported incidence of yersiniosis. The decline was interrupted in 1998, and since then the incidence has been between 50 and 150 notified cases per year.

Studies conducted during the 1980s revealed that a large proportion of Norwegian pigs were carriers of *Y. enterocolitica* serogroup O:3 and that the same variant frequently could be isolated from pig carcasses. In 1995-1996 a serological survey of all multiplier herds (n=66) belonging to the cooperative slaughterhouse organisation showed that 35.5% of the fattening pigs had antibodies against *Y. enterocolitica* O:3, and 80% of the herds had at least one pig (of 40 pigs tested per herd) with antibodies against *Y. enterocolitica* O:3. In another survey where blood samples from 5 fatteners in each of 326 randomly selected herds were analysed for antibodies against *Y. enterocolitica* O:3, 53% of the pigs and 64% of the herds tested positive.

In 1997-1998, 300 samples of raw pork products were analyzed. *Y. enterocolitica* O:3 was isolated from 2% of the samples by a culturing method (NMKL method no. 117), while use of a PCR method indicated the presence of pathogenic *Y. enterocolitica* in 17% of the samples. This was lower than in a similar survey conducted in 1988-1989.

National evaluation of the recent situation, the trends and sources of infection

From 1994 to 1998, a reduction in the incidence of yersiniosis in humans was observed. This decline coincided with a gradual introduction of improved slaughter routines with the aim of preventing pig carcasses from becoming contaminated with *Y. enterocolitica*.

Relevance of the findings in animals, feedingstuffs and foodstuffs to human cases

Pork products are generally considered the most important source of yersiniosis in humans. A Norwegian case control study conducted in the period 1988-1990 identified consumption of such products as an important risk factor in addition to consumption of untreated drinking water and a general preference for undercooked meat. In 2006 two smaller outbreaks of yersiniosis both linked to a traditional cold cuts pork product were reported.

Recent actions taken to control the zoonoses

During the mid 1990s, there was a gradual introduction of improved slaughter routines that aid in preventing pig carcasses from being contaminated with *Y. enterocolitica*. A significant reduction of reported cases of human yersiniosis cases was noted parallel to this.

Yersiniosis in humans

Reporting system in place for the human cases

Human cases are reported to the Norwegian Surveillance System for Communicable Diseases (MSIS), from microbiological laboratories as well as from clinical doctors. The system distinguishes between domestic and imported cases. The severity of the disease at the time of reporting is also recorded. However, the surveillance system does not follow individual patients over time to record further disease development and final outcome. Cases confirmed by serology only are also reported, but due to recent changes in laboratory practices these are not included in this report.

Case definition

A case from which *Y. enterocolitica* or *Y. pseudotuberculosis* has been isolated or a clinical compatible case with an epidemiological link to a culture confirmed case.

Diagnostic/analytical methods used

Bacteriology (isolation of *Yersinia* species) followed by voluntary confirmation (species identification and serotyping) at the National Reference Laboratory.

Notification system in place

According to the Communicable Disease Act, human cases are notifiable to the Norwegian Surveillance System for Communicable Diseases (MSIS) since 1992.

History of the disease and/or infection in the country

In the years 1982-1994, the number of notified cases varied between 154 and 274 (mean 187, median 182). From 1994 there was a steady decline in yersiniosis reports. This decline coincided with a gradual introduction of improved routines when slaughtering pigs, which resulted in reduced contamination with *Y. enterocolitica* to pig carcasses. The decline was interrupted in 1998, and since then the incidence has been between 50 and 150 notified cases per year.

Results of the investigation

In 2010, a total of 52 cases of yersiniosis were reported (incidence rate 1.1 per 100 000). A total of 25 (48%) cases were domestic.

National evaluation of the recent situation, the trends and sources of infection

Although the incidence of yersiniosis has decreased in recent years and the number of registered cases is moderate, the disease is still the fourth most commonly recorded foodborne zoonotic infection in Norway. Moreover, the majority of the cases have acquired the infection within Norway. The vast majority of cases are sporadic. The most common serogroup is O:3. The number of cases reported in 2010 is almost the same as in 2008, which had the lowest number of reported cases since the surveillance of yersiniosis started.

Relevance as zoonotic disease

Yersiniosis is an important zoonotic disease in Norway, with the majority of cases acquired within Norway. Pigs are considered to be a major reservoir, and pork products are considered to be an important source for pathogenic *Y. enterocolitica*, although uncertainties still remain regarding the epidemiology.

Additional information

Patients whose work represents a risk for spread of the disease, e.g., in food production and health care, are advised to stay away from work as long as they have symptoms. It is recommended that for these patients two consecutive faecal samples examined after the symptoms have disappeared should be negative before returning to work.

Yersinia in animals

Monitoring system

There are no official monitoring programmes for *Y. enterocolitica* in live animals or in animals at slaughter.

Recent actions taken to control the zoonoses

During the mid 1990s, there was a gradual introduction of improved slaughter routines that aid in preventing pig carcasses from being contaminated with *Y. enterocolitica*. A significant reduction in the incidence of reported yersiniosis in humans was noted subsequent to this action.

Measures in case of the positive findings or single cases

None.

Results of the investigation

In 2010, a total of one wild hare was diagnosed with *Y. pseudotuberculosis*.

Trichinellosis

General evaluation of the national situation

History of the disease and/or infection in the country

Trichinellosis has been found sporadically in farmed food producing animals and was last detected in two pig herds in 1994. This was the first report of trichinellosis in pigs since 1981. Trichinellosis occurs endemically among wild red foxes in mainland Norway and among wild arctic foxes and polar bears in the archipelago of Svalbard. In a survey in red foxes killed during the licensed hunting season in 1994-1995 and 2002-2005, 4.8% of 393 examined animals were positive for *Trichinella* larvae. *T. spiralis* and *T. pseudospiralis* were not found in these studies. *T. nativa* is the most commonly found species in Norwegian foxes. Trichinellosis has also been diagnosed in farmed foxes. Human trichinellosis acquired in Norway has not been reported since 1980. The two last reported cases of human trichinellosis, in 1996, were both imported.

National evaluation of the recent situation, the trends and sources of infection

Trichinellosis was last detected in food producing animals in 1994, in two pig herds. Trichinellosis occurs endemically among wildlife.

Relevance of the findings in animals, feedingstuffs and foodstuffs to human cases

As Norwegian food producing animals very rarely are infected with *Trichinella*, and all slaughtered pigs and horses are analysed for the parasite, the probability of contracting trichinellosis from food producing animals of Norwegian origin is close to zero.

Trichinellosis in humans

Reporting system in place for the human cases

Human cases are reported to the Norwegian Surveillance System for Communicable Diseases (MSIS), from microbiological laboratories as well as from clinical doctors. The system distinguishes between domestic and imported cases. The severity of the disease at the time of reporting is also recorded. However, the surveillance system does not follow individual patients over time to record further disease development and final outcome.

Case definition

A clinically compatible case that is laboratory confirmed.

Diagnostic/analytical methods used

Muscle biopsy and histopathology (demonstration of *Trichinella* larvae in tissue) and serology.

Notification system in place

According to the Communicable Disease Act, human cases are notifiable to the Norwegian Surveillance System for Communicable Diseases (MSIS) since 1975.

History of the disease and/or infection in the country

Human trichinellosis acquired in Norway is very rare, the last case being reported in 1980. The last two cases of imported trichinellosis were reported in 1996, in immigrants from ex-Yugoslavia.

Results of the investigation

In 2010, no cases of human trichinellosis were reported.

Relevance as zoonotic disease

The risk of acquiring trichinellosis from domestic sources is considered very low because trichinellosis only has been detected twice in food producing animals since 1981, extensive surveillance programmes are in place, and the production is intensive and takes place under controlled conditions.

Additional information

If a human case should be diagnosed, epidemiological investigations will be initiated in order to identify the source and prevent further cases.

Trichinella in animals

A. *Trichinella* in pigs

Monitoring system

All pigs must be controlled for *Trichinella* at slaughter according to Council Directive 77/96/EEC (in force until 1st March) and Regulation (EC) No 2075/2005 (as from 1st March 2010).

Frequency of the sampling

Every slaughtered animal is sampled.

Type of specimen taken

Diaphragm muscle.

Methods of sampling (description of sampling techniques)

Methods used are in accordance to Council Directive 77/96/EEC (still in force until 1st March 2010) and Commission regulation (EC) No 2075/2005 of 5 December 2005 laying down specific rules on official controls for *Trichinella* in meat. Up to 100 samples, each of 1 gram, can be analysed as a pooled sample when using a digestion method. Occasionally, the trichinoscopic method is used instead of a digestion method.

Case definition

An animal with a positive test result in the official examination.

Diagnostic/analytical methods used

Artificial digestion method of pooled samples. Occasionally a compression (trichinoscopic) method is used.

Preventive measures in place

It is prohibited to feed pigs with swills. Most pig herds have implemented programs for combating of rodents (rats and mice).

Control program/mechanisms

All pigs must be controlled for *Trichinella* at slaughter according to Council Directive 77/96/EEC until 1st March 2010 and Regulation (EC) No 2075/2005 thereafter.

Measures in case of the positive findings or single cases

Measures are imposed on holdings with positive findings of *Trichinella* in accordance with Regulations concerning measures against contagious animal diseases of 27.06.2002 no 732 (movement restrictions, epidemiological investigation into the source and possible spread of the disease). Detection of *Trichinella* must be reported immediately. Farms delivering positive carcasses will be identified. Animals from such farms will be given special attention at slaughter the following six months. The sample size for the digestion method will be increased to 2 grams.

Notification system in place

Trichinellosis has been a notifiable disease (List B) since 1965.

Results of the investigation

In 2010, no cases of trichinellosis among slaughtered pigs were reported.

National evaluation of the recent situation, the trends and sources of infection

Trichinellosis was last detected in two pig herds in 1994.

Relevance of the findings in animals to findings in foodstuffs and to human cases

There have not been any findings of *Trichinella* in pigs or pig meat for many years. The risk of obtaining trichinellosis from Norwegian pig meat is negligible.

B. *Trichinella* in horses

Monitoring system

All horses were controlled for *Trichinella* at slaughter in accordance with Council Directive 77/96/EEC until March 2010 and according to Regulation (EC) No 2075/2005 since 1st March 2010.

Frequency of the sampling

Every slaughtered animal is sampled.

Type of specimen taken

Tongue or masseter muscle.

Methods of sampling (description of sampling techniques)

Methods used are in accordance with Council Directive 77/96/EEC and Regulation (EC) No 2075/2005 since 1st March 2010. For analyses, 5 g per animal is included in a pooled sample of maximum 100 g.

Case definition

An animal with a positive test result in the official examination.

Diagnostic/analytical methods used

Artificial digestion method of pooled samples.

Results of the investigation

In 2010, no cases of trichinellosis were reported among slaughtered horses.

Measures in case of the positive findings or single cases

All horse carcasses that are included in a positive pooled sample will be retested individually (samplesize 5 and 50 grams respectively). In accordance with Regulation No 732 of 27 June 2002 on measures against contagious animal diseases measures, such as movement restrictions and investigations into the source of the disease and any spread, are imposed on holdings with positive findings of *Trichinella*. Detection of *Trichinella* must be reported immediately.

Notification system in place

Trichinellosis has been a notifiable List B disease since 1965.

Relevance of the findings in animals to findings in foodstuffs and to human cases

There have not been any findings of *Trichinella* in horses or horse meat. The risk of obtaining trichinellosis from Norwegian horse meat is negligible.

C. *Trichinella* spp. in wild animals

Monitoring system

All wild boars and bears must be controlled for *Trichinella* at slaughter in accordance with Council Directive 77/96/EEC and (in force at the beginning of 2010) and Regulation (EC) No 2075/2005 since 1st March 2010. This control is compulsory. Wild and farmed foxes and other species of wildlife are occasionally sampled.

Frequency of the sampling

Depending on the situation and animal species.

Type of specimen taken

Diaphragm, tongue, masseter or occasionally other muscles.

Methods of sampling (description of sampling techniques)

Depending on the situation and animal species.

Case definition

An animal with a positive test result.

Diagnostic/analytical methods used

Digestion methods or trichinoscopic (compression) method.

Measures in case of the positive findings or single cases

If trichinellosis is diagnosed in a farmed fox, the animal holding will get official restrictions in accordance with Regulations concerning measures against contagious diseases of 27.06.2002 no 732 (not allowed to sell animals and epidemiological investigation of the source and any spread of the infection).

Notification system in place

Trichinellosis has been a notifiable disease since 1965.

Results of the investigation

In 2010, two foxes (*Vulpes vulpes*) were investigated and found negative for *Trichinella*.

National evaluation of the recent situation, the trends and sources of infection

Trichinellosis occurs endemically among wildlife.

Echinococcosis

General evaluation of the national situation

History of the disease and/or infection in the country

E. granulosus used to be relatively common in semi-domesticated reindeer in North Norway until the 1950s (approx. 10% prevalence in the 1950s). Today the parasite has virtually been eliminated as a result of systematic anthelmintic treatment of herder dogs and a reduction in the feeding of raw offal from slaughter to the herder dogs. In 2003, one reindeer had pathological findings compatible with *E. granulosus* infestation. *E. granulosus* was last reported in cattle in 1987.

E. multilocularis has never been detected in mainland Norway in any animal species. In 1999, a research project on echinococcosis in the archipelago of Svalbard detected *E. multilocularis* cysts in the liver of 16% of 172 sibling voles tested. In a follow-up study, faecal samples from polar foxes, dogs, and cats were collected. The parasite was diagnosed in three of six polar foxes, in one of 48 dogs, and in neither of the two cats. The methods used were coproantigen ELISA, egg isolation and PCR. The number of voles that annually tested positive between 2000-2006, varied between 19% and 96%. In mainland Norway in the period 2002-2005, a total of 314 red foxes were investigated using coproantigen ELISA, egg isolation and PCR, all were negative for *E. multilocularis*. An ongoing national surveillance program for *E. multilocularis* was implemented in red foxes in 2006. Since then, 1305 foxes have been examined using egg isolation and PCR. All of the total 1619 red foxes have tested negative for *E. multilocularis*.

Human echinococcosis has never been a public health problem in Norway.

National evaluation of the recent situation, the trends and sources of infection

The risk of acquiring echinococcosis in Norway is considered very low. The pathological finding compatible with *E. granulosus* infestation in a reindeer in 2003 is a reminder that this parasite still may be present and that this requires awareness in the reindeer industry, especially with regard to the importance of regular treatment of herd dogs with an anthelmintic drug. The occurrence of *E. multilocularis* among animals in the archipelago of Svalbard requires alertness among health personnel, especially in this region.

Relevance of the findings in animals, feedingstuffs and foodstuffs to human cases

The pathological finding compatible with *E. granulosus* infestation in a reindeer in 2003 is a reminder that this parasite still may be present and that this requires awareness in the reindeer industry. As *E. multilocularis* has never been detected in mainland Norway in any animal species, the risk to humans of contracting *E. multilocularis* infection in mainland Norway is probably very low. The occurrence of *E. multilocularis* among animals in the archipelago of Svalbard requires alertness among health personnel in this region. Inhabitants of Svalbard have been informed about the risk.

Echinococcosis in humans

Reporting system in place for the human cases

Human cases are reported to the Norwegian Surveillance System for Communicable Diseases (MSIS), from microbiological laboratories as well as from clinical doctors. The system distinguishes between domestic and imported cases. The severity of the disease at the time of reporting is also recorded. The surveillance system does not follow individual patients over time to record further disease development and final outcome.

Case definition

A clinical compatible case that is laboratory confirmed.

Diagnostic/analytical methods used

Serology and histopathology.

Notification system in place

According to the Communicable Disease Act, human cases are notifiable to the Norwegian Surveillance System for Communicable Diseases (MSIS) since 1 July 2003.

History of the disease and/or infection in the country

Human echinococcosis has never been a public health problem in Norway and the incidence is considered to be at most very low. The majority of cases have acquired the infection abroad.

Results of the investigation

In 2010, one case of infection with *E. granulosus* was reported. The patient was infected abroad.

Relevance as zoonotic disease

The risk of acquiring echinococcosis in Norway is considered very low. The pathological finding compatible with *E. granulosus* infestation in a reindeer in 2003 is a reminder that this parasite still is around and that this requires awareness in the reindeer industry, especially as regard the importance of regular treatment of herd dogs with an antihelminthic drug. As *E. multilocularis* has never been detected in mainland Norway in any animal species, the risk to humans of contracting echinococcosis caused by *E. multilocularis* in mainland Norway is close to zero. The presence of *E. multilocularis* among animals in the archipelago of Svalbard requires vigilance amongst health personnel in this region. Inhabitants of Svalbard have been informed about the risk.

Echinococcus in animals

A. *E. granulosus* in animals

Monitoring system

Surveillance in intermediate hosts is achieved through the official meat inspection. There are no official monitoring programmes for *Echinococcus granulosus* among the final hosts (dogs).

Frequency of the sampling

All possible intermediate hosts subject to meat inspection procedure according to Council Directive 64/433/EEC (still in force January and February 2010) and Regulation (EC) No 2075/2005 since 1st March 2010. An exception to this is wild ruminants in which there is no obligatory control if they are shot for private consumption.

Methods of sampling (description of sampling techniques)

Inspection for hydatid cysts at the abattoir.

Case definition

An animal with a positive test result.

Diagnostic/analytical methods used

Macroscopic (visual) examination of organs and parasitology.

Other preventive measures than vaccination in place

Dogs and cats imported to Norway, except those imported from Sweden and Finland, must be treated with an anthelmintic drug the last ten days before entering Norway and again one week after arrival. Treatment with an anthelmintic drug is also advocated on a general basis, especially for herd dogs in areas with reindeer.

Control program/mechanisms

Mandatory official meat control.

Measures in case of the positive findings or single cases

An animal with cystic echinococcosis will be condemned. Epidemiological data will be collected in order to find the source of infection and measures will be introduced to prevent further spread.

Notification system in place

Echinococcosis has been a notifiable List B disease according to the Animal Diseases Act since 1985.

Results of the investigation

In 2010, all slaughtered animals subjected to official meat control were negative for *E. granulosus*. No cases of infection with *E. granulosus* were diagnosed in carnivores.

Additional information

Methods in use when examining final hosts: Faecal material: Egg isolation (flotation) and PCR.

B. *E. multilocularis* in animals

Monitoring system

In 2006 a National surveillance programme regarding *E. multilocularis* in red foxes was started. The program also included the examination of samples from about 300 hunted foxes collected for parasitological research purposes in the period 2002-2005. In 2010, no animals were investigated. Animals hunted late 2010 will be investigated in 2011 together with animals hunted in the spring 2011. There are no official monitoring programmes for *E. multilocularis* in other animals.

Methods of sampling (description of sampling techniques)

Foxes: Faecal samples.

Case definition

An animal with a positive test result.

Diagnostic/analytical methods used

Faecal samples: Taeniid egg isolation and multiplex PCR techniques.

Other preventive measures than vaccination in place

Dogs and cats imported to Norway, except those imported from Sweden and Finland, must be treated with an anti-helminthic drug the last ten days before entering Norway and also one week after arrival. Treatment with an anti-helminthic drug is also advocated on a general basis. Due to findings of *E. multilocularis* in the archipelago of Svalbard, the Norwegian Food Safety Authority requires that dogs and cats that are introduced into mainland Norway from Svalbard must be treated with an anti-helminthic drug approved for treatment of *E. multilocularis*.

Recent actions taken to control the zoonoses

The findings of *E. multilocularis* in the archipelago of Svalbard in 1999 resulted in follow-up studies, requirements regarding anti-helminthic treatment of dogs and cats in regard to export, and an information campaign directed to the inhabitants of Svalbard.

Notification system in place

Echinococcosis has been a notifiable List B disease according to the Animal Diseases Act since 1985.

Results of the investigation

In 2010, one dog and one racoon dog (*Nyctereutes procyonoides*) were investigated. Both were negative.

National evaluation of the recent situation, the trends and sources of infection

In mainland Norway, *E. multilocularis* has never been detected in any animal species. The main host of *E. multilocularis*, the red fox, has been investigated by examining a total of 1633 foxes killed during hunting from 2002-2009. All foxes have been negative. Thus, there are so far no indications that this parasite has established in Norway. In 1999, in a research project on echinococcosis in the archipelago of Svalbard, *E. multilocularis* was detected in 16% of 172 sibling voles tested. In a follow-up study, the parasite was diagnosed in samples from polar foxes and one dog. Of the voles tested in 2000-2006, between 19% and 96% were positive each year.

Toxoplasmosis

General evaluation of the national situation

History of the disease and/or infection in the country

In 1994, the last year human toxoplasmosis was notifiable, 33 cases were reported (incidence rate 0.77 per 100 000 inhabitants) of which eight were children less than one year. *Toxoplasma gondii* is endemic in animals in Norway with the domestic cat and wild lynx being the final hosts. Studies indicate that the parasite is relatively common among sheep; 18% of the lambs were seropositive in a survey conducted during the 1990s, and seropositive lambs were identified on 44% of the farms included. The parasite is assumed to be less common among Norwegian pigs. In the above mentioned survey, 2% of the slaughtering pigs tested were seropositive. In 2008, a survey using goat sera collected in the period 2002-2008 were tested. A total of 18.5% of the animals were positive. Also wild ruminants (cervids) can be infected; a survey carried out among 4300 cervids killed during hunting in 1992-2000, revealed 34% seropositive roe deer, 13% seropositive moose, 8% seropositive red deer and 1% seropositive reindeer.

National evaluation of the recent situation, the trends and sources of infection

Toxoplasma gondii is endemic in Norway with the domestic cat and wild lynx being the final hosts. Studies indicate that the parasite is relatively common among sheep and goat and less common among Norwegian pigs. Also wild ruminants (cervids) can be infected. There are no data indicating recent developments in the prevalence of the infection in various species.

Relevance of the findings in animals, feedingstuffs and foodstuffs to human cases

A case control study designed to identify risk factors for maternal toxoplasma infection during pregnancy showed that the following exposures were associated with an increased risk: Eating raw or undercooked minced meat, eating unwashed raw vegetables or fruits, eating raw or undercooked mutton, eating raw or undercooked pork, cleaning the cat litter box and washing the kitchen knife infrequently after preparing raw meat. This implies that Norwegian farm animals and food products of Norwegian origin may well be an important source of human toxoplasmosis.

Toxoplasmosis in humans

Reporting system in place for the human cases

Human cases are not reported to the Norwegian Surveillance System for Communicable Diseases (MSIS).

Case definition

A clinically compatible case that is laboratory confirmed.

Diagnostic/analytical methods used

Serology (antibody detection) and parasitological examination (identification of parasite in clinical specimens).

Notification system in place

Since 1995, human toxoplasmosis has not been a notifiable disease in Norway.

History of the disease and/or infection in the country

In different epidemiological surveys conducted in Norway, 7-27% of pregnant women tested have been seropositive. The percentages have been age dependent, with the proportion of seropositive individuals increasing with age, and have also varied with region and ethnicity. It is estimated that approximately 90% of fertile women are susceptible to the disease and that approximately two out of 1000 susceptible pregnant women are infected during pregnancy. In 1994, the last year human toxoplasmosis was notifiable, 33 cases were reported (incidence rate 0.77 per 100 000 inhabitants) of which eight were children less than one year.

Results of the investigation

The disease is not notifiable.

National evaluation of the recent situation, the trends and sources of infection

Toxoplasma gondii is endemic in Norway although the parasite is considered to be somewhat less prevalent as compared to countries more south in Europe. The public health importance of toxoplasmosis is its potential of causing severe disease in infants who are born to women infected during pregnancy, and its potential of causing severe disease in immunocompromised individuals, such as people with AIDS. Seroprevalence surveys among pregnant women indicate that infection with *Toxoplasma* is common in Norway. Pregnant women are advised how to avoid infection during pregnancy.

Relevance as zoonotic disease

A case control study designed to identify risk factors for maternal toxoplasma infection during pregnancy showed that the following exposures were associated with an increased risk: Eating raw or undercooked minced meat, eating unwashed raw vegetables or fruits, eating raw or undercooked mutton, eating raw or undercooked pork, cleaning the cat litter box and washing the kitchen knife infrequently after preparing raw meat. This implies that Norwegian farm animals and food products of Norwegian origin may well be an important source of *Toxoplasma* for spread to humans.

Toxoplasma in animals

Monitoring system

Sampling of animals is performed in case of clinical suspicion and in connection to import/export. Surveys are occasionally performed.

Frequency of the sampling

In cases of clinical suspicion or specific surveys.

Case definition

An animal with a positive test result.

Diagnostic/analytical methods used

Serology (direct agglutination test) or pathology.

Measures in case of the positive findings or single cases

Normally none.

Notification system in place

Toxoplasmosis in animals has been a List C disease according to the Animal Diseases Act since 1965.

Results of the investigation

In 2010, several animal species were investigated for *Toxoplasma* at the Norwegian Veterinary Institute. Only for sheep were there more than five investigated animals and at least one positive animal: A total of 23 out of 49 investigated sheep were positive.

National evaluation of the recent situation, the trends and sources of infection

Toxoplasma gondii is endemic in Norway. There are no data indicating recent developments in the prevalence of the infection in various species.

Relevance of the findings in animals to findings in foodstuffs and to human cases

A risk for humans of contracting toxoplasmosis in Norway does exist. However, the relevance of clinical toxoplasmosis is most important in immunosuppressed persons and in pregnant women.

Rabies

General evaluation of the national situation

History of the disease and/or infection in the country

Rabies in animals has not been recorded in mainland Norway. An epidemic occurred in the arctic fox population in the archipelago of Svalbard in 1980, with diagnosed cases also in reindeer and one seal. Since then, sporadic cases have been diagnosed in arctic foxes, the last case in 1999. During the period 1980 - 2009, 25 animal cases were diagnosed with this disease. However, transmission of rabies to humans has never been recorded in the archipelago of Svalbard.

National evaluation of the recent situation, the trends and sources of infection

The favourable situation in mainland Norway regarding rabies is stable. However, there are concerns about the risk of introducing rabies through illegally imported dogs.

Relevance of the findings in animals, feedingstuffs and foodstuffs to human cases

Rabies has sporadically been diagnosed in wild animals in the archipelago of Svalbard, the last occurrence was in 1999. Although no transmission of rabies to humans has been recorded in Svalbard, people being in contact with wild animals in Svalbard should be aware of the risk and vaccination is recommended. In mainland Norway, the possible introduction of rabies through illegally imported animals is a concern.

Rabies in humans

Reporting system in place for the human cases

Human cases are reported to the Norwegian Surveillance System for Communicable Diseases (MSIS), from microbiological laboratories as well as from clinical doctors. The system distinguishes between domestic and imported cases. The severity of the disease at the time of reporting is also recorded. However, the surveillance system does not follow individual patients over time to record further disease development and final outcome. Cases are also reported immediately to the Municipal Medical Officer. If a domestic animal source is suspected, the Municipal Medical Officer also informs the Norwegian Food Safety Authority. Investigations will be initiated in order to identify the source and prevent further cases.

Case definition

A clinical case that is laboratory confirmed.

Diagnostic/analytical methods used

Detection of viral antigens by an immunofluorescence test in neurological tissue (usually brain) in connection to post-mortem examination, virus isolation in cell culture, or identification of an antibody titre greater than the threshold value in serum or cerebrospinal fluid from an unvaccinated person.

Notification system in place

According to the Communicable Disease Act, human cases are notifiable to the Norwegian Surveillance System for Communicable Diseases (MSIS) since 1975.

History of the disease and/or infection in the country

Human rabies was last described in Norway in 1815.

Results of the investigation

In 2010, no human cases were reported.

Relevance as zoonotic disease

As mainland Norway has been free from rabies for almost two centuries and stringent regulation regarding import of animals are in place, the risk of contracting rabies in mainland Norway is close to zero. Rabies has sporadically been diagnosed in wild animals in the archipelago of Svalbard, the last time in a fox found dead in 1999. Although no transmission of rabies to humans has been recorded in Svalbard, people being in contact with wild animals in Svalbard should be aware of the risk.

Additional information

Rabies vaccine containing inactivated virus is available for the following indications: Pre-exposure prophylaxis to; 1) individuals with prolonged travels to countries with high incidence of rabies; 2) individuals who will work with animals in endemic areas; 3) persons who are at frequent risk of bites from bats; 4) laboratory personnel involved in rabies diagnostics. Post-exposure prophylaxis to individuals presumably exposed to rabies virus abroad or in the archipelago of Svalbard, or who have been bitten by bats. The post-exposure prophylaxis includes specific antiserum in addition to the vaccine.

Lyssavirus (rabies) in animals

A. Rabies in dogs

Monitoring system

There are no active surveillance programmes regarding rabies. However, being a notifiable disease, clinical suspicion of rabies must be reported immediately.

Frequency of the sampling

On clinical suspicion.

Type of specimen taken

Brain.

Methods of sampling (description of sampling techniques)

The brain is removed at autopsy, and samples are taken according to the procedures described in the OIE manual.

Case definition

A case that is laboratory confirmed.

Diagnostic/analytical methods used

The Fluorescent antibody test (FAT) is the OIE prescribed test for rabiesvirus antigen and is performed according to the OIE Terrestrial Manual 2010. In addition, molecular methods (real time RT-PCR, RT-PCR and gene sequencing) are used.

Vaccination policy

Vaccines containing inactivated rabies virus antigen are available for dog, cat and ferret. Vaccination is required for international transport of these animal species in compliance with national regulations. For dogs living in Svalbard vaccination is a mandatory requirement. Otherwise, vaccination against rabies is not done on a routine basis in mainland Norway.

Other preventive measures than vaccination in place

Infected animals will be destroyed and measures taken to prevent further cases.

Control program/mechanisms

Dogs, cats and ferrets entering Norway from countries not considered rabies free (include all third countries not listed in the annex "Listed third countries according to Regulation (EC) No 988/2003"), are subject to four months of quarantine in an officially approved station, followed by a two months period in home quarantine. However, dogs, cats and ferrets from EEA countries not considered rabies free are permitted into Norway without quarantine, provided they have been vaccinated against rabies and have been proven antibody positive according to a given protocol.

Measures in case of the positive findings or single cases

Infected animals will be destroyed and measures taken to prevent further cases.

Notification system in place

Rabies has been a notifiable List A disease according to the Animal Diseases Act since 1965. Rabies is dealt with in Council Directive 92/65/EEC, which is implemented in Regulations on animal health conditions regarding movements, import and export of certain animals [FOR 2004-07-01 No 1105 and FOR 2004-02-20 No 464].

Results of the investigation

In 2010 no cases were reported. Two dogs were investigated and found negative.

National evaluation of the recent situation, the trends and sources of infection

Mainland Norway is recognized as rabies free. However, there are concerns regarding a possible increase in the number of illegally imported dogs. Rabies has sporadically been diagnosed in wild animals in the archipelago of Svalbard, the last time in a fox found dead in 1999. Although no transmission of rabies to dogs has been recorded in Svalbard, owners must respect and follow up the mandatory vaccination programme.

B. Rabies virus in wildlife

Monitoring system

There are no active surveillance programmes regarding rabies. However, the disease must be reported immediately on clinical suspicion.

Frequency of the sampling

On clinical suspicion. In Svalbard, dead foxes and other animals should be secured for laboratory examination.

Type of specimen taken

Brain, in bats also oral swabs.

Methods of sampling (description of sampling techniques)

The brain is removed at autopsy, and samples are taken according to the procedures described in the OIE manual.

Case definition

A case that is laboratory confirmed.

Diagnostic/analytical methods used

The Fluorescent antibody test (FAT) is the OIE prescribed test for rabiesvirus antigen and is performed according to the OIE terrestrial Manual 2010. In addition, molecular methods (real time RT_PCR, RT-PCR and gene sequencing) are used.

Measures in case of the positive findings or single cases

Infected animals will be destroyed and measures taken to prevent further cases.

Notification system in place

Rabies has been a notifiable List A disease according to the Animal Diseases Act since 1965. Rabies is dealt with in Council Directive 92/65/EEC, which is implemented in Regulations on animal health conditions regarding movement, import and export of certain animals [FOR 2004-07-01 No 1105 and FOR 2004-02-20 No 464].

Results of the investigation

In 2010, nine wild animals were tested and all were found negative. The majority of animals came from the Svalbard area (seven arctic foxes (*Vulpes lagopus*) and one polar bear (*Ursus maritimus*)). In addition one red fox (*Vulpes vulpes*) came from mainland Norway.

National evaluation of the recent situation, the trends and sources of infection

Mainland Norway is considered rabies free. Rabies has sporadically been diagnosed in wild animals in the archipelago of Svalbard, the last time in a fox found dead in 1999. Although no transmission of rabies to other animal species has been recorded in Svalbard, people in Svalbard should be aware of the risk.

Q-fever

General evaluation of the national situation

History of the disease and/or infection in the country

Q-fever has not been diagnosed in animals in Norway. In a survey in 2008, bulk milk samples from 460 dairy herds and 550 blood samples from 55 suckling cattle herds were sampled in five cattle dense counties (Rogaland, Trøndelag, Hedmark, Oppland and Østfold). In surveys performed in 2009, samples from 349 goat herds (mainly bulk milk samples from 2005-2009), samples from 121 sheep herds and 45 cattle herds were analysed for Q-fever. All samples were negative.

National evaluation of the recent situation, the trends and sources of infection

C. burnetii has never been detected in animals in Norway.

C. burnetii in animals

Sampling strategy

Surveys are performed occasionally.

Case definition

Sample positive for antibodies against *C. burnetii*.

Diagnostic/analytical methods used

Detection of antibodies to *C. burnetii* in milk or serum by ELISA.

Results of the investigation

In 2010, a total of 3420 cattle from 3378 herds (mainly bulk milk samples), 49 sheep (from one herd), 15 swine (from a breeding company - export control) and 102 alpaca (from 5 imported herds) were tested for Q-fever. All samples were negative.

Information on specific indicators of antimicrobial resistance

Escherichia coli, non-pathogenic

National evaluation of the recent situation, the trends and sources of infection

Earlier surveys as well as data from the monitoring programme NORM-VET indicate a low to moderate prevalence of resistance in indicator *E. coli* from Norwegian food producing animals and food. Those resistances that are most commonly encountered are to antimicrobials that have been or still are typically used therapeutically such as streptomycin, sulphonamides, tetracycline and ampicillin. Fluoroquinolone resistance is rarely detected, which is a reflection of a very low use of such antimicrobials in food producing animals in Norway.

Sampling strategy used in monitoring

The sampling of animals for isolation of indicator *E. coli* to be included in resistance monitoring is a part of the Norwegian monitoring programme for antimicrobial resistance in feed, food and animals, NORM-VET. The sampling is spread throughout the year and each year one or several animal species are included. In 2010, cattle and wild red fox were monitored.

Type of specimen taken

Faecal material.

Methods of sampling (description of sampling techniques)

The samples were taken as part of other surveillance programmes.

Procedures for the selection of isolates for antimicrobial testing

Only one isolate from each herd (cattle) or animal (fox) was included.

Methods used for collecting data

All samples were sent to the Norwegian Veterinary Institute in Oslo for identification and for antimicrobial susceptibility testing.

Laboratory methodology used for identification of the microbial isolates

A sample was plated directly onto the surface of lactose-saccharose-bromthymol blue agar without broth enrichment. After incubation of the agar plates at 37°C for 24 h, a typical colony was plated onto blood agar (Heart infusion agar (Difco) containing 5% bovine blood) and incubated at 37°C for 18-24 h. Colonies were identified as *E. coli* by typical appearance, lactose and/or saccharose fermentation and a positive indole reaction.

Laboratory methods used for detection for resistance

The VetMIC microdilution method (Dept. of Antibiotics, National Veterinary Institute, Sweden) was used for the susceptibility testing of all isolates. The antimicrobials included are listed in the table.

Breakpoints used in testing

For interpretation of results epidemiological cut-off values recommended by EFSA were applied. When no cut-off value was recommended, a cut-off value was defined on basis of the actual MIC distributions obtained in the NORM-VET programme.

Control program/mechanisms

The sampling of animals for isolation of indicator *E. coli* to be included in resistance monitoring is a part of the Norwegian monitoring programme for antimicrobial resistance in feed, food and animals, NORM-VET.

Results

The qualitative data are presented in the table. Quantitative data as well as data on breakpoints and range of testing are presented in the NORM/NORM-VET 2010 report.

Table Antimicrobial susceptibility testing of *E. coli*

	Cattle	Fox
Number of isolates tested	209	55
Percentage of isolates resistant to		
Tetracycline	1.9	1.8
Chloramphenicol	0	0
Cefotaxime	0.5	0
Nalidixic acid	0	1.8
Sulfamethoxazole	3.3	5.5
Trimethoprim	0	1.8
Streptomycin	9.1	3.6
Gentamicin	0	0
Ampicillin	1.9	1.8

Information on specific microbiological agents

Histamine in foodstuffs

Monitoring system

Regular testing of selected species is required as an internal part of food business operator's quality assurance system. Surveys are performed occasionally.

Definition of positive finding

Histamine values above 100 mg/kg.

Diagnostic/analytical methods used

Reverse phase HPLC/UV

Table Histamine in foodstuffs

	Source of information	Sampling unit	Sample weight	Units tested	Total units in non-conformity	<=100 mg/kg	>100 - <=200 mg/kg	>200 - <=400 mg/kg	>400 mg/kg
Fish	NIFES	single	5 g	25	0	25*			

* All samples <5.5 mg/kg

Foodborne outbreaks

Foodborne outbreaks are incidences of two or more human cases of the same disease or infection where the cases are linked or are probably linked to the same food source. Situation, in which the observed human cases exceed the expected number of cases and where a same food source is suspected, is also indicative of a foodborne outbreak.

System in place for identification, epidemiological investigations and reporting of foodborne outbreaks

Health personnel are required to report suspected foodborne outbreaks to the Municipal Health Officer, who is required to report to the County Governor (Fylkesmannen) and to the Norwegian Institute of Public Health. Suspected outbreaks are reported immediately to the Municipal Medical Officer who notifies the Norwegian Institute of Public Health the same day. If a domestic food or animal source is suspected, the Municipal Medical Officer also informs the local Food Safety Authority.

The Norwegian Food Safety Authority has voluntary reporting where the District Offices report foodborne outbreaks.

Norway has since 2005 a web-based reporting system called Vesuv where all outbreaks in humans are to be reported and stored in a database at the Norwegian Institute of Public Health. Vesuv came in a new version in 2010.

If an indigenous outbreak is suspected, epidemiological investigations will be initiated in order to identify the source and prevent further cases. For imported cases, the country of acquisition will be recorded. If information through international networks indicates that a case belongs to an outbreak, epidemiological investigations will be initiated.

Description of the types of outbreaks covered by the reporting

All suspected foodborne outbreaks are notifiable. The definition of a foodborne outbreak is two or more human cases with the same infection where the cases are linked or are probably linked to the same food source, or when observed number of human cases exceeds the expected number of cases during the same time period and place, and food is a likely vehicle.

Trends in numbers of outbreaks and numbers of human cases involved

The number of reported foodborne outbreaks has increased in Norway since the web-based reporting system was established in 2005 (42 in 2005, 65 in 2006 and 80 in 2007). We believe that this increasing trend is due to a higher reporting frequency rather than a real higher number of outbreaks. The number of reported outbreaks decreased again in 2008 (64), 2009 (47) and was stable in 2010 (53).

Relevance of the different causative agents, food categories and the agent/food category combinations

Traditionally, the most common cause of foodborne outbreaks in Norway has been bacterial intoxication (*Clostridium perfringens*, *Bacillus cereus* and *Staphylococcus aureus*). Recently, foodborne outbreaks of norovirus caused by infected food handlers and imported food items have become more common. Reported domestic outbreaks of salmonellosis and campylobacteriosis have been relatively rare.

Relevance of the different type of places of food production and preparation in outbreaks

Traditionally, outbreaks have mainly been associated with inadequate handling and temperature abuse, causing food intoxication. In addition, untreated water has caused several outbreaks.

Evaluation of the severity and clinical picture of the human cases

In 2010, one severe outbreak was reported. No deaths were related to foodborne outbreaks.

Descriptions of single outbreaks of special interest

We registered one severe outbreak caused by sorbitol fermenting *E. coli* O157 including three children and all of them developed haemolytic uremic syndrome (HUS). The source of the outbreak was not found.

Table Reported foodborne outbreaks in Norway 2010 - summarized data

Agent	Total number of outbreaks (verified outbreaks)	Human cases	Implicated food in verified outbreaks
<i>Bacillus</i>	2	5	
<i>Campylobacter</i>	5	18	
<i>E. coli</i> , pathogenic	1	3	
Foodborne viruses	24 (4)	588	Raw oyster (2), Lollo lettuce, Mixed salad
Other agents ¹	3	11	
<i>Salmonella</i>	3	25	
Unknown	15	139	

¹ Hepatitis A, *Francisella tularensis*, Staphylococcal enterotoxins