

The surveillance programme for *Echinococcus multilocularis* in red foxes (Vulpes vulpes) in Norway 2022



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Summary

The prevalence of *Echinococcus multilocularis* was based on PCR analysis of faecal samples from 503 red foxes (*Vulpes vulpes*) collected during the licensed fox-hunting season in 2022 and 24 grey wolves (*Canis lupus*) killed in 2022. None of the samples tested positive for *E. multilocularis*, documenting that the prevalence in carnivore hosts (foxes and wolves) were below 1% at a confidence level of at least 95%.

Introduction

Echinococcus multilocularis (Figure 1), the parasite causing alveolar echinococcosis in humans, is endemic in several regions of the northern hemisphere, including eastern and central parts of Europe (1, 2). During the past decades, the prevalence of *E. multilocularis* in Europe has increased in the known endemic areas (3), and the geographic distribution has expanded to regions where the parasite appeared to be absent previously (4). Similarly, alveolar echinococcosis, the life-threatening zoonotic disease caused by the metacestode stage of this tapeworm, is increasing in prevalence in Europe. A recent European project ranked *E. multilocularis* first amongst the food-borne parasites based on public health concerns (5).

The adult tapeworm resides in the small intestine of wild carnivores (definitive hosts) such as red foxes, raccoon dogs and wolves. Domestic dogs and cats can also act as definitive hosts if they prey on infected small mammals, predominantly rodents that serve as intermediate hosts.



Figure 1: Echinococcus multilocularis, adult worm used for spiking of positive controls included in the PCR analyses. The sack-like uterus containing hundreds of eggs is clearly visible. Worms used as controls were inactivated by kept frozen for <75 C for several days, and subsequently stored in 70% ethanol. Professor Peter Deplazes, University of Zurich, kindly donated the depicted worm. Photo: Øivind Øines, Norwegian Veterinary Institute

In Scandinavia, the first discovery of *E. multilocularis* was on the high-arctic Norwegian islands of Svalbard (6) and in Denmark (7) in 1999. However, there was no evidence of its presence in mainland Fennoscandia (8) until its detection in Sweden in February 2011 (9). Despite analyses of more than 8100 faecal samples from foxes since 2002 (10, present report), *E. multilocularis* has not been reported in mainland Norway.

Anthelmintic treatment of dogs, prior to import from endemic regions, is compulsory in Norway to prevent introduction of the parasite. According to the EU Directive 576/2013 and Commission Delegated Regulation (EU) 2018/772 on pet movement, the maintenance of this national regulation requires the documentation of an *E. multilocularis*-free status within the country in question. The results of the investigations in the surveillance programme to EFSA every year to document freedom of *Echinococcus multilocularis* in mainland Norway. Every year the datasets from participating countries is subject to assessment and the resulting report; "Annual assessment of *Echinococcus multilocularis* surveillance reports submitted in 20xx in the context of Commission Delegated Regulation (EU) 2018/772" is published in the EFSA journal.

The Norwegian Food Safety Authority (NFSA) is responsible for implementing the surveillance programme. The Norwegian Veterinary Institute (NVI) is responsible for sampling plans, laboratory investigations and the reporting components of the programme.

Aim

The aim of the surveillance is to document the freedom of *E. multilocularis* in mainland Norway.

Materials and methods

In the *E. multilocularis* surveillance of 2022, faecal samples collected from red foxes (*Vulpes* vulpes) hunted during the licensed hunting season (i.e. January to mid-April and mid-July to late December 2022) were included. In addition to faeces from foxes, samples from wolves (*Canis lupus*) killed legally, or illegally during 2022, were tested for the presence of *E. multilocularis*.

The RiBESS tool (https://shiny-efsa.openanalytics.eu/app/ribess) was used to estimate the sample size required to substantiate the absence of the parasite from the target population with a confidence level of 95%. For the calculation we used sensitivity value 0.63, specificity value 1.00 (11), together with an estimated population size of 151000 red foxes.



Figure 2: Map showing observations of red fox in Norway This is an online service where citizens can logon and register their observations of fauna and flora in Norway. Source: Norwegian Biodiversity Information Centre <u>https://artsdatabanken.no/Pages/180936/Roedrev</u> Eide NE (2015). Rødrev Vulpes vulpes (Linnaeus, 1758). <u>www.artsdatabanken.no/Pages/180936. Downloaded</u> <u>26/05/2022</u>.

Recruitment of hunters was done through the webpages of the Norwegian Veterinary Institute. The hunters enter their name and municipality via the webpages of the Norwegian Veterinary Institute (https://www.vetinst.no/nyheter/registrering-som-provetaker-av-rodrev). This registration is announced on NVI's web page and at the NVI's Facebook page. Those that have contributed to the program previous years are invited by e-mail to register, but the registration is also open for new hunters. The selection of foxhunters has then been based on residence and previous quality of their submitted samples. In addition, the selection also includes some hunters that are new to the programme and therefore covers some new regions.

Sampling containers and detailed instructions for sampling were sent to the hunters who were selected for participation in the program. The samples were submitted to the laboratory with written information on sample locality, date of the sampling, sex (male or female) and estimated age of the animal (juvenile or adult) in pre-paid envelopes. All counties in Norway were included in the sampling regime.



The majority of samples were collected in January, February and March (Fig. 3).

Figure 3: Temporal distribution of samples (Source: EFSA). In Norway red fox hunting is allowed all year except between 15th April - 15th July.

Individual faecal samples (3 g per animal) were analysed using the sensitive DNA-fishing (magnetic capture) method combined with real-time PCR detection of *E. multilocularis* mtDNA. This procedure involves magnetic capture of biotin tagged DNA-hybridisation probes targeting a locus on the *E. multilocularis* mtDNA. The biotin attached to the hybridisation probe/target DNA-complex is coupled by a noncovalent protein-protein binding interaction to streptavidin molecules which are coated onto magnetic beads. This allow extraction of parasite mtDNA from inhibitors and other DNA in the sample, by using a magnet (11).

Detection of the *E. multilocularis* DNA was carried out by real-time PCR (11, 12). If a positive real-time PCR signal is detected, the presence of *E. multilocularis* mtDNA can be verified by an additional independent real-time PCR (12), and /or using a standard PCR targeting the nad1 gene followed by Sanger-sequencing (13). All tests are performed in duplicates with each run including two positive control DNA samples (from adult worms) and negative controls (MilliQ water) included in each run.

The DNA-fishing method is capable of detecting *E. multilocularis* DNA originating from any developmental stage of the parasite, including worms, and eggs in high volume samples. The method is suitable for use during the patent phase of the infection when eggs are shed in the faeces. This period constitutes roughly two-thirds of the entire infection period. The MC-DNA/realtime PCR methods has been shown to be more sensitive than egg isolation by sieving followed by detection of parasite DNA using a multiplex PCR, used previously in the Norwegian surveillance program (11, 12).

Initially, a test sensitivity of 63% and a specificity of 100% were assumed 11). However, our internal validation has demonstrated an overall sensitivity of 0.80. For samples spiked with \geq 10 eggs the sensitivity is 0.90 which is close to the method of Isaksson *et al.* (12) (Se 0.88) (11, 14). The apparent prevalence and corresponding confidence interval were estimated using Epitools (15), with a test sensitivity of 0.63 and a specificity of 1.00.

Results and Discussion

In 2022, 527 faecal samples from wild carnivores were analysed for *E. multilocularis*: 503 samples from red foxes (Table 1, Figure 4) and 24 samples from wolves (*Canis lupus*) (Table 1, Figure 5). All samples tested negative for *E. multilocularis* giving an estimated apparent prevalence of 0% (0.0 - 0.7%, 95%CI).

Surveillance results were no different from earlier years. All faecal samples collected from wild carnivores in mainland Norway as part of the surveillance program in 2022, were negative by PCR for *E. multilocularis*.

According to requirements of Regulation (EU) No 2018/772, Annex I, the disease freedom status must have a pathogen-specific surveillance program designed to detect a prevalence of $\leq 1\%$ at minimum confidence level of 95%. The number of samples collected and analysed in Norway in 2022 was sufficient to document a current prevalence of *E. multilocularis* below 1%.

Table 1: Number and origin (county) of red foxes and wolves examined for Echinococcus multilocularis in Norway
during the red fox licensed hunting season in 2022 (January to mid-April and mid-July to late December) and
corresponding numbers for the period 2002 - 2021.

County 2022	County 2019	Number of	Other species	
		2022	Total 2002-2022	tested 2022
Viken	Østfold	78	1 001	1
	Akershus	56	817	2
	Buskerud	27	444	
Oslo	Oslo	9	188	
Innlandet	Hedmark	43	1 128	20
	Oppland	20	539	1
Vestfold og Telemark	Vestfold	2	127	
	Telemark	34	373	
Agder	Aust-Agder	38	176	
	Vest-Agder	5	165	
Rogaland	Rogaland	12	145	
Vestland	Hordaland	39	382	
	Sogn og Fjordane	23	314	
Møre og Romsdal	Møre og Romsdal	22	219	
Trøndelag	Trøndelag	27	1 049	
Nordland	Nordland	26	357	
Troms og Finnmark	Troms	37	548	
	Finnmark	5	176	
Total	Total	503	8 148	24 wolves

However, it is worrying that the rising prevalence in countries close to Norway has increased the risk of introduction of the parasite to Norway. In Sweden, there are already detections of *E. multilocularis* in four different regions (10, 21), and surveillance in Denmark has demonstrated its presence in two regions (16). Studies in Sweden have discovered *E. multilocularis* in the intermediate hosts of field vole (*Microtus agrestis*) and water voles (*Arvicola amphibious*) in two study areas (20). Moreover, studies in the Baltics have shown a wider distribution of the tapeworm than previously anticipated, which has caused an increasing number of alveolar echinococcosis cases in humans (17). This is worrying, as a lack of compliance with the anthelmintic treatment requirements for pets entering the Norway after having visited endemic areas has been demonstrated (18, 19). The above-mentioned points illustrate why it is imperative to continue with the surveillance for *E. multilocularis* in Norway to document and ensure Norway has a continuous disease-free status via the annual surveillance program.

Our results support the continuing national regulation for compulsory anthelmintic treatment of imported dogs to minimize the risk of an introduction of *E. multilocularis* to Norway.



Figure 4: Map of Norway showing the origin of red foxes by municipality, tested for Echinococcus multilocularis during the red fox licensed hunting season for red fox in 2022.



Figure 5: Map of Norway showing the origin of wolfs by municipality, tested for Echinococcus multilocularis in 2022

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