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The surveillance programme for Gyrodactylus salaris in Atlantic salmon and rainbow trout in Norway 2025

REPORT 18/2026

The surveillance programme for *Gyrodactylus salaris* in Atlantic salmon and rainbow trout in Norway 2025

Authors

Haakon Hansen, Gunn Jorid Fornes, Saima Nasrin Mohammad, Steinar Gilleberg Stensli, Mari Berger Skjøstad, Hilde Irene Welde, David Allan Strand and Marit Måsøy Amundsen

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Content

Summary 3

Introduction 3

Aims 4

Materials and methods 4

Results and discussion 6

Acknowledgements 6

References 7

Appendix A 8

Appendix B 10

Summary

In 2025, a total of 2367 specimens of Atlantic salmon from 71 rivers and 2340 specimens of Atlantic salmon and rainbow trout from 71 hatcheries/farms were sampled and examined in the surveillance program for *Gyrodactylus salaris*. In addition, five farms rearing rainbow trout and two hatcheries rearing Arctic char, were surveyed using eDNA monitoring. The parasite was not detected in any of these samples.

A total of 54 rivers have so far been confirmed infected since *G. salaris* was introduced to Norway. As of 31.12.25, *G. salaris* is confirmed present in six Norwegian river systems in the Drammen infection region. These are the rivers Drammenselva, Lierelva, Sande, Selvikvassdraget, Bergerelva and Ebbestadelva. Treatment of five rivers in the Driva infection region, Batnfjordselva, Driva, Litledalselva, Usma and Gylelva in county Møre og Romsdal, was completed in 2024. These rivers are at present part of the declaration of freedom programme, awaiting confirmation of eradication. Altogether 43 rivers have been declared free after treatment.

Introduction

In the period from 1975 until the start of 2025, pathogenic strains of *Gyrodactylus salaris* had been detected on Atlantic salmon (*Salmo salar*) fingerlings/parr in 54 rivers, 13 hatcheries/farms with Atlantic salmon parr/smolts and 26 hatcheries/farms with rainbow trout (*Oncorhynchus mykiss*). In addition, both pathogenic and non-pathogenic strains of *G. salaris* have been found on Arctic char (*Salvelinus alpinus*).

The policy of the Norwegian authorities is to eradicate *G. salaris* from infected watersheds and farms (Anon, 2014). If *G. salaris* is detected in a farm, eradication is carried out by eliminating the hosts (Atlantic salmon and/or rainbow trout). This also ensures elimination of the parasite since it lacks specialised free-living stages and does not use intermediate hosts in its life cycle. In rivers, the eradication is done by chemical treatment. In most instances rotenone has been the preferred chemical, but one exception to this is the treatment of River Lærdalselva in 2011-2012, where acidified aluminium sulphate was used to eradicate the parasite (Hindar et al., 2015). Recently, a full-scale treatment using chlorine as the main chemical has been completed in river Driva, Møre og Romsdal county (Garvik et al., 2025). In contrast to rotenone treatment, treatment with aluminum sulfate and chlorine will kill the parasite, but not the host.

By the entrance to 2025, *G. salaris* was confirmed eradicated from 43 rivers and from all hatcheries/fish farms. I *Gyrodactylus salaris* was still present in 6 river systems namely Drammenselva (012.Z), Lierelva (011.Z), Ebbestadelva (012.2Z) in county Buskerud, Vesleelva (Sandeelva)(013.Z) and Selvikvassdraget (013.1Z) in county Vestfold. In addition Batnfjordselva (108.3Z), Driva (109.Z), Litledalselva (109.5Z), Usma (109.4Z) and Gylelva (109.7Z) in county Møre og Romsdal are at present part of the declaration of freedom programme, awaiting confirmation of eradication.

Gyrodactylus salaris is included in the list F of nationally listed and notifiable diseases, and Norway has implemented national measures for the parasite which comply with Regulation (EU) 2016/429, article 226 (3). *Gyrodactylus salaris* is also listed as a notifiable aquatic animal disease by the World Organization for Animal Health (WOAH). Surveillance for *G. salaris*, aiming to declare freedom from the parasite in treated rivers, has been ongoing since the early 1980s. The Norwegian Veterinary Institute (NVI) coordinates the surveillance programme on behalf on the Norwegian Food Safety Authority (NFSA) and publishes the overall results in annual reports available on the NVI website (www.vetinst.no).

NFSA is responsible for the sampling in fish farms. NVI is responsible for the sampling in the rivers, but County Environmental Departments and other institutions/companies are commissioned to do the actual sampling. NVI

is responsible for examination of the fish samples and the subsequent species identification, if *Gyrodactylus* is detected.

Aims

The surveillance program aims to document the absence of *G. salaris* in Norwegian farms and rivers, as well as to detect and track any spread of the parasite to new river systems or fish farms.

Materials and methods

The selection of rivers for inclusion in the surveillance programme follow specified criteria which takes into account the risk of infection with *G. salaris* (see text box 1). In general, a total of 30 wild Atlantic salmon juveniles are sampled from each selected river, preferably from three different sites located far apart. To increase the sensitivity of the surveillance for the River Rana (Nordland county), where the source of the infection detected in 2014 remains unknown, an additional sample of 30 fish is taken one month after the first sample. In Tana (Troms and Finnmark county), 150 salmon are sampled from 15 sites due to the large size of this watercourse. Fingerlings/parr/smolts of an age of 1+ or older (preferred size ranging from 7 - 12 cm) are caught by means of electrofishing. The fish are killed and then preserved whole in 96% ethanol.

In farms and hatcheries, either 30 Atlantic salmon or 60 rainbow trout are sampled by seine net. The fish are killed and all fins (except the adipose fin) are cut off and preserved in 96% ethanol. Each farm/hatchery is examined every second year.

All samples are sent to the NVI for examination under a stereo microscope at 10 - 15 times magnification. For wild Atlantic salmon, the whole surface of the fish, including the skin, head, fins and gills, is examined, while only the fins from farmed fish are examined.

When *Gyrodactylus* specimens are detected, species determination is performed by NVI. NVI is the WOAH reference laboratory for "Infection with *Gyrodactylus salaris*" and the methods used for species identification follow those given by the WOAH Manual of Diagnostic Tests for Aquatic Animals: https://www.woah.org/fileadmin/Home/eng/Health_standards/aahm/current/2.3.03_G_salaris.pdf

In addition to the standard surveillance methods where fins of fish are examined under a stereo microscope, environmental DNA (eDNA) monitoring was applied for the first time in this surveillance program, and the method was applied to survey two hatcheries rearing Arctic char and five farms rearing rainbow trout. This is also the first time hatcheries rearing Arctic char were included in this surveillance program. It is important to include this species in the surveillance as some populations of Arctic char have been shown to be suitable hosts for G. salaris (Mo et al., 2023; Robertsen et al., 2007).

Environmental DNA monitoring is a tool that can detect minute amounts of DNA in water samples using a combination of water filtering and molecular detection. All organisms in water shed cells containing DNA into the environment (Thomsen et al., 2012). By using species-specific primers and probes and sensitive PCR-methods, it is possible to detect and identify the presence of DNA from specifically targeted species in water samples. This method is also developed for detecting *G. salaris* (Rusch et al., 2018) and has previously been applied in field studies in the River Drammenselva and elsewhere (Fossøy et al., 2019; Hytterød et al., 2018; Rusch et al., 2018).

In addition to the two hatcheries with Arctic char, five farms rearing rainbow trout in the county Innlandet were also monitored using eDNA methodology (see Table 1 and Appendix B for details). The low number and/or high economic value of each fish in these farms made such non-destructive monitoring especially attractive.

From all hatcheries/farms, four samples were taken: one from the water intake, one from the outlet and the two last from different locations within the farm. Water samples of 1-2 litres were collected and filtered on site onto Dual eDNA filters (0.45 µm pore size, Sylphium, Groningen, The Netherlands) by pumping the water through the filters manually using a 50 ml syringe. Qiagen ATL buffer was added and samples were shipped to NVI for analyses.DNA was isolated in the laboratory using a Nucleospin Plant II midi kit and Qiagen buffer according to Fossøy et al. [15].

The eDNA samples from the farms with rainbow trout were analysed with qPCR assays designed to detect *G. salaris* (Rusch et al., 2018), and rainbow trout (Wilcox et al., 2015). The assay for rainbow trout is included as positive control as large amounts of DNA from the host should be present in the water and thus on the filters and should amplify in the qPCR-analyses. At the time of the analyses an assay for Arctic char was not available at NVI and thus, eDNA samples from the Arctic char hatcheries were thus analysed only with qPCR assays designed to detect *G. salaris* (Rusch et al., 2018).

The qPCR analyses on the eDNA samples were run with three technical replicates. The Cq cut-off value was set to 40, and each sample were considered positive if two out of the three replicates had a Cq value below 40.

Criteria for inclusion of rivers in the surveillance program for *Gyrodactylus salaris* in short*.

1. Rivers declared free from infection after treatment. This criteria states that when a watercourse is declared free from infection with *G. salaris*, it should be included in the surveillance program for a minimum of five (5) years. After five years, a watercourse can be excluded from the surveillance program unless it fulfils other risk factors for their continued inclusion (see below).

2. Large salmon rivers in terms of spawning targets. This criteria states that the 30 largest salmon rivers in terms of spawning targets should be included in the surveillance program.

3. Rivers with a high risk of inter-river dispersal of *G. salaris*. This criteria states that rivers with a high risk of being infected via inter-river (brackish-water) dispersal of *G. salaris* should be included in the surveillance program. Due to the decreasing numbers of infected rivers in Norway, the number of rivers included based on fulfilment of this criteria has decreased, and will continue to decrease, when further rivers are declared free from infection.

4. Rivers with other risk of infection: this criteria overlaps somewhat with criteria 3, but the main focus is on the threat from areas bordering other countries.

5. Geographic coverage: This criteria states that a minimum of two (2) rivers from each county where salmon rivers are present should be included in the surveillance program.

*For further details please consult the following document: Reply from the Norwegian Veterinary Institute (NVI) to the Norwegian Food Safety Authority (NFSA) 5th February 2020: FSA reference number 2020/173134, alt. NVI reference number 20/12419.

Results and discussion

Altogether, 2367 specimens of Atlantic salmon and one (1) Arctic char from 71 rivers and 2340 specimens of Atlantic salmon and rainbow trout from 71 farms were examined in 2025 (See Table 1 and appendix A and B for details). In addition, eDNA monitoring was used to survey two hatcheries with Arctic char and five farms with rainbow trout. Buskerud is not represented as its rivers are part of the Drammen infection region and excluded from the surveillance programme. *Gyrodactylus salaris* was not detected in any of the samples examined in the surveillance programme.

The status at the end of the year 2025 is thus that *G. salaris* is present in six Norwegian river systems. All six rivers; Drammenselva, Lierelva, Sande, Selvikvassdraget, Bergerelva and Ebbestadelva, are part of the Drammen infection region. Treatment of five rivers in the Driva infection region was completed in 2024, and these rivers are at present part of the declaration of freedom programme, awaiting confirmation of eradication.

Table 1. Number of rivers, farms and fish examined for *Gyrodactylus salaris* in 2025 (See Appendix A and B for details).

County	n rivers	species *	n examined	positive	n farms*	species	n examined	positive
Innlandet	-	-	-	-	2	RT	120	0
Østfold	2	AS	60	0	-	-	-	-
Oslo	1	AS	30	0	-	-	-	-
Akershus	4	AS	115	0	-	-	-	-
Buskerud	-	-	-	-	-	-	-	-
Vestfold	2	AS	63	0	-	-	-	-
Telemark	2	AS	63	0	2	AS/RT	90	0
Agder	4	AS	122	0	1	AS	33	0
Rogaland	3	AS	93	0	5	AS	145	0
Vestland	5	AS	154	0	22	AS/RT	783	0
Møre og Romsdal	8	AS	206	0	11	AS	329	0
Trøndelag	10	AS	309	0	13	AS/RT	390	0
Nordland	10	AS	368	0	8	AS	248	0
Troms	6	AS	188	0	6	AS	172	0
Finnmark	14	AS	596	0	1	AS	30	0
Total	**71**		**2367**	**0**	**71**		**2340**	**0**

[1]AS = Atlantic salmon, RT = rainbow trout. *In addition, two hatcheries rearing Arctic char, *Salvelinus alpinus*, and five farms rearing rainbow trout, *Oncorhynchus mykiss*, all in County Innlandet, were analysed by eDNA monitoring (See text and Appendix B for details).

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Appendix A

Watercourses examined for Gyrodactylus salaris in 2024 sorted by watercourse code. D = Detected, ND = Not detected.

County	Watercourse	Watercourse code	No of Atlantic salmon examined	*G. salaris*
Østfold	Enningdalselva	001.1Z	30	ND
Østfold	Glommavassdraget	002.Z	30	ND
Akershus	Hølenelva	004.Z	33	ND
Akershus	Årungselva	005.3Z	19	ND
Oslo	Lysakerelva	007.Z	30	ND
Akershus	Sandvikselva	008.Z	33	ND
Akershus	Årosvassdraget	009.Z	30	ND
Vestfold	Aulivassdraget	014.Z	33	ND
Vestfold	Numedalslågen	015.Z	30	ND
Telemark	Herreelva	016.4Z	30	ND
Telemark	Skiensvassdraget	016.Z	33	ND
Agder	Arendalsvassdraget	019.Z	30	ND
Agder	Tovdalselva	020.Z	31	ND
Agder	Otra	021.Z	30	ND
Agder	Mandalselva	022.Z	31	ND
Rogaland	Bjerkreimselva	027.Z	31	ND
Rogaland	Figgjo	028.Z	31	ND
Rogaland	Suldalslågen	036.Z	31	ND
Vestland	Vossovassdraget	062.Z	32	ND
Vestland	Lærdalselva	073.Z	31	ND
Vestland	Sogndalselva	077.3Z	31	ND
Vestland	Gaula	083.Z	30	ND
Vestland	Nausta	084.7Z	30	ND
Møre og Romsdal	Måna	103.1Z	33	ND
Møre og Romsdal	Innfjordselva	103.2Z	30	ND
Møre og Romsdal	Breidvikelva	103.42Z	12	ND
Møre og Romsdal	Isa	103.4Z	30	ND
Møre og Romsdal	Skorgeelva	103.5Z	20	ND
Møre og Romsdal	Rauma	103.Z	30	ND
Møre og Romsdal	Flemma	109.13Z	21	ND
Møre og Romsdal	Surna	112.Z	30	ND
Trøndelag	Orkla	121.Z	32	ND
Trøndelag	Gaula	122.Z	32	ND
Trøndelag	Nidelvvassdraget	123.Z	30	ND
Trøndelag	Stjørdalsvassdraget	124.Z	32	ND
Trøndelag	Verdalsvassdraget	127.Z	31	ND
Trøndelag	Figga/Snåsavassdraget	128.3Z	31	ND
Trøndelag	Steinkjervassdraget	128.Z	30	ND
Trøndelag	Stordalselva	135.Z	30	ND

Trøndelag	Årgårdsvassdraget	138.Z	30	ND
Trøndelag	Namsen	139.Z	31	ND
Nordland	Hestdalselva	149.61Z	28	ND
Nordland	Halsanelva	149.6Z	30	ND
Nordland	Hundåla	151.1Z	30	ND
Nordland	Vefsna	151.Z	31	ND
Nordland	Drevja	152.2Z	30	ND
Nordland	Fusta	152.Z	60	ND
Nordland	Leirelva	153.22Z	30	ND
Nordland	Røssåga	155.Z	30	ND
Nordland	Ranavassdraget	156.Z	69	ND
Nordland	Saltdalsvassdraget	163.Z	30	ND
Troms	Salangselva	191.Z	30	ND
Troms	Nordkjoselva	198.Z	31	ND
Troms	Signaldalelva	204.Z	34	ND
Troms	Skibotnvassdraget	205.Z	32	ND
Troms	Manndalselva	206.1Z	32	ND
Troms	Reisavassdraget	208.Z	29	ND
Finnmark	Altavassdraget	212.Z	32	ND
Finnmark	Repparfjordvassdraget	213.Z	31	ND
Finnmark	Stabburselva	223.Z	35	ND
Finnmark	Lakselvvassdraget	224.Z	30	ND
Finnmark	Børselvvassdraget	225.Z	31	ND
Finnmark	Storelva	228.Z	30	ND
Finnmark	Tana	234.Z	179	ND
Finnmark	Komagelva	239.Z	31	ND
Finnmark	Vestre jakobselv	240.Z	34	ND
Finnmark	Vesterelva	241.5Z	31	ND
Finnmark	Munkelva	244.4Z	31	ND
Finnmark	Neidenvassdraget	244.Z	36	ND
Finnmark	Karpelva	247.3Z	33	ND
Finnmark	Grense Jakobselv	247.Z	32	ND

[3]In addition to the Atlantic salmon, 1 Arctic char, *Salvelinus alpinus*, from Ytterbekken, a small stream draining into the same estuary as Ranavassdraget and with a previous history of infection were also sampled and found negative. This specimen is not included in the total in this table. River Målselva (water course code 196.Z) and River Pasvik (water course code 246.Z) were originally included in the surveillance for 2025 but were excluded due to too high water level/bad weather at the time of sampling.

Appendix B

Farms and hatcheries examined for Gyrodactylus salaris in 2024 grouped by county from south to north. AS= Atlantic salmon, RT= Rainbow trout, AC=Arctic char, eDNA = environmental DNA monitoring, ND = Not detected, NA = Not Available

County	Farm/Hatchery	Hatchery code	Fish species	No. of AS/RT examined	*G. salaris*
Innlandet	Begna (Henrik Haadem)	12517	RT	eDNA	ND
Innlandet	Fasle (Henrik Haadem)	33977	RT	60	ND
Innlandet	Ferisfjorden	13881	RT	60	ND
Innlandet	Lofoss Mølle	12342	RT	eDNA	ND
Innlandet	Lomen Slidrefjorden	12341	RT	eDNA	ND
Innlandet	Løpet	12545	AC	eDNA	ND
Innlandet	Nedre Hande	13716	RT	eDNA	ND
Innlandet	Noråker Gård	10364	RT	eDNA	ND
Innlandet	Snerta	Cultivation	AC	eDNA	ND
Telemark	Kjølebrønn	12961	AS	30	ND
Telemark	Rjukan I	36957	RT	60	ND
Agder	Fjellsæ I	10581	AS	33	ND
Rogaland	Fister	10123	AS	30	ND
Rogaland	Klekkjeriet på Ritland	(tom)	AS	30	ND
Rogaland	Lerangsvågen Land	11927	AS	29	ND
Rogaland	Tytlandsvik	35857	AS	26	ND
Rogaland	Vågafossen	11892	AS	30	ND
Vestland	Arnafjord	12173	RT	61	ND
Vestland	Barlindbotn	13843	AS	30	ND
Vestland	Bjølvefossen	12172	AS	30	ND
Vestland	Eidesvik	11606	AS	30	ND
Vestland	Femangervågen	12112	AS	30	ND
Vestland	Fjæra	12073	AS	30	ND
Vestland	Flatraker	13826	RT	60	ND
Vestland	Gjæravågen	11589	AS	30	ND
Vestland	Gjølanger	11795	AS	30	ND
Vestland	Herand	13157	AS	30	ND
Vestland	Hermansverk	12165	RT	62	ND
Vestland	Kjærelva	11493	AS	30	ND
Vestland	Kvernhusvika	13327	AS	30	ND
Vestland	Ljonesvågen	12079	AS	30	ND
Vestland	Ljøsne klekkeri	12343	AS	30	ND
Vestland	Sagvåg	28216	AS	30	ND
Vestland	Skogseidevatnet	12042	AS	30	ND
Vestland	Skålevik	11540	AS	30	ND
Vestland	Strømsnes	11648	RT	60	ND
Vestland	Sævareid	10141	AS	30	ND
Vestland	Ænes	12084	AS	30	ND
Vestland	Øyerhamn	12032	AS	30	ND

Møre og Romsdal	Aunvågen	10221	AS	30	ND
Møre og Romsdal	Barstadvik	10183	AS	30	ND
Møre og Romsdal	Dale	12217	AS	30	ND
Møre og Romsdal	Haukvik Kraft - smolt stamfisk	Cultivation	AS	30	ND
Møre og Romsdal	Hjelvik	13672	AS	30	ND
Møre og Romsdal	Kjørsvikbugen	12415	AS	29	ND
Møre og Romsdal	Moltustranda	12325	AS	30	ND
Møre og Romsdal	Steinsvik	12222	AS	30	ND
Møre og Romsdal	Sætre	13671	AS	30	ND
Møre og Romsdal	Urke	12269	AS	30	ND
Møre og Romsdal	Videild	12223	AS	30	ND
Trøndelag	Beslvik	13964	AS	30	ND
Trøndelag	Bessaker	12596	AS	30	ND
Trøndelag	Follafoss	13958	AS	30	ND
Trøndelag	Kvernavikvatnet	12686	AS	30	ND
Trøndelag	Lennavika	13742	AS	30	ND
Trøndelag	Lensvik	13179	AS	30	ND
Trøndelag	Moldtua	12737	AS	30	ND
Trøndelag	Nernesset	13178	AS	30	ND
Trøndelag	Olden	12745	AS	30	ND
Trøndelag	Sunnskjør	23735	AS	30	ND
Trøndelag	Survik	12672	AS	28	ND
Trøndelag	Årvika	10406	AS	30	ND
Trøndelag	Mosvik Klekkeri (Slira)	Cultivation	AS	32	ND
Nordland	Bjørnsøya	13185	AS	30	ND
Nordland	Breivika	13811	AS	30	ND
Nordland	Forsan	33217	AS	30	ND
Nordland	Framnes	10496	AS	30	ND
Nordland	Glomfjord II	24016	AS	40	ND
Nordland	Mastermovika II	15315	AS	30	ND
Nordland	Mo Industripark	11064	AS	28	ND
Nordland	Reppen	34097	AS	30	ND
Troms	Hellaren	11335	AS	30	ND
Troms	Jøvik	11333	AS	30	ND
Troms	Salangsverket	36357	AS	30	ND
Troms	Sandøra	36477	AS	30	ND
Troms	Sørfjorden	13946	AS	20	ND
Troms	Vinterneset	39737	AS	32	ND
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